

17 β -Estradiol, a potential ally to alleviate SARS-CoV-2 infection

Ana Cristina Breithaupt-Faloppa ^{1,*} Cristiano de Jesus Correia ¹ Carla Máximo Prado ^{II} Roberta Sessa Stilhano ^{III} Rodrigo Portes Ureshino ^{IV,V} Luiz Felipe Pinho Moreira ^I

^ILaboratório de Cirurgia Cardiovascular e Fisiopatologia da Circulação (LIM-11), Instituto do Coração (InCor), Faculdade de Medicina FMUSP, Universidade de São Paulo, São Paulo, SP, BR. ^{II}Instituto de Saúde e Sociedade (ISS), Universidade Federal de São Paulo (UNIFESP), Santos, SP, BR. ^{III}Faculdade de Ciências Médicas da Santa Casa de São Paulo (FCMSCSP), São Paulo, SP, BR. ^{IV}Departamento de Ciências Biológicas, Universidade Federal de São Paulo (UNIFESP), Diadema, SP, BR. ^VLaboratório de Endocrinologia Molecular e Translacional, Escola Paulista de Medicina, Universidade Federal de São Paulo (UNIFESP), São Paulo, SP, BR.

Breithaupt-Faloppa AC, Correia CJ, Prado CM, Stilhano RS, Ureshino RP, Moreira LFP. 17 β -Estradiol, a potential ally to alleviate SARS-CoV-2 infection. *Clinics*. 2020;75:e1980

*Corresponding author. E-mail: ana.breithaupt@hc.fm.usp.br

Considering that female sexual hormones may modulate the inflammatory response and also exhibit direct effects on the cells of the immune system, herein, we intend to discuss the sex differences and the role of estradiol in modulating the lung and systemic inflammatory response, focusing on its possible application as a treatment modality for SARS-CoV-2 patients. COVID-19 patients develop severe hypoxemia early in the course of the disease, which is silent most of the time. Small fibrinous thrombi in pulmonary arterioles and a tumefaction of endothelial were observed in the autopsies of fatal COVID-19 cases. Studies showed that the viral infection induces a vascular process in the lung, which included vasodilation and endothelial dysfunction. Further, the proportions of CD4+ T and CD8+ T lymphocytes were strongly reduced in patients with severe SARS-CoV-2 infection. Estradiol is connected with CD4+ T cell numbers and increases T-reg cell populations, affecting immune responses to infection. It is known that estradiol exerts a protective effect on endothelial function, activating the generation of nitric oxide (NO) via endothelial nitric oxide synthase. Estrogen attenuates the vasoconstrictor response to various stimuli and induces vasodilation in the pulmonary vasculature during stress situations like hypoxia. It exerts a variety of rapid actions, which are initiated after its coupling with membrane receptors, which in turn, may positively modulate vascular responses in pulmonary disease and help to maintain microvascular flow. Direct and indirect mechanisms underlying the effects of estradiol were investigated, and the results point to a possible protective effect of estradiol against COVID-19, indicating that it may be considered as an adjuvant therapeutic element for the treatment of patients affected by the novel coronavirus.

KEYWORDS: 17 β -Estradiol; COVID-19; Infection; Coagulation; Inflammation.

■ INTRODUCTION

Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2), a novel coronavirus, is responsible for the pandemic viral disease outbreak that originated in Wuhan, China, in December 2019. The disease is characterized by severe acute respiratory syndrome, and the virus has already infected more than 3 million people worldwide. No therapies have been shown to be effective to date (1). Data regarding the clinical management of the disease have been published, some of which have demonstrated that there are differences in prevalence and mortality regarding sex (2-4). There is a higher predominance of men affected in several countries (5);

in Italy, a prevalence of 70% of men among deceased people was reported (6). Recent data reported by the New York City Department of Health and Mental Hygiene (7) confirmed the higher prevalence and mortality of the disease in men. Moreover, a study of nine pregnant women infected with SARS-CoV-2 showed that the infection did not evolve to severe coronavirus disease 2019 (COVID-19) during the pregnancy (8).

Although there is evidence that the immune response (innate and adaptive) differs between men and women, sexual dimorphism does not receive deserved attention as a potential factor for understanding the different immune responses observed between men and women (9,10). In fact, clinical studies show that women have a lower incidence of pneumonia, sepsis, and multiple organ failure than men (11-13). There are several reports on the participation of female sex hormones in mediating the repercussions caused by trauma followed by hemorrhagic shock (14-23). Important differences exist in epidemiology, pathophysiology, and treatment of cardiovascular diseases, such as coronary artery disease, hypertension, cardiomyopathy, and heart failure. Cardiovascular disorders are also more severe in postmenopausal women (24).

Copyright © 2020 CLINICS – This is an Open Access article distributed under the terms of the Creative Commons License (<http://creativecommons.org/licenses/by/4.0/>) which permits unrestricted use, distribution, and reproduction in any medium or format, provided the original work is properly cited.

No potential conflict of interest was reported.

Received for publication on May 5, 2020. **Accepted for publication on** May 5, 2020

DOI: 10.6061/clinics/2020/e1980



Suba (25) revealed that epidemiological data from SARS and MERS CoV epidemics indicated that the outcome of human coronavirus infections is strongly sex-dependent, suggesting the role of a strong protective estrogen signal in adult female patients compared to age-matched males. In this context, considering that female sexual hormones may modulate the inflammatory response and also exhibit direct effects on the cells of the immune system, here we intend to discuss the sex differences and the role of estradiol in modulating the lung and the systemic inflammatory response and its possible application as a treatment for SARS-CoV-2 patients.

The effects of SARS-CoV-2

The inflammatory response is a natural defense mechanism of the body to remove harmful stimuli such as pathogens and initiate the recovery process. SARS-CoV-2, the new coronavirus that induces COVID-19, enters the body through the interaction between the S protein on the virus surface and angiotensin-converting enzyme-2 (ACE-2) molecules expressed in epithelial cells in the lungs (26). After SARS-CoV-2 replication in the respiratory and intestinal epithelium cells (27), it can induce an immune response that causes several alterations in the lung including inflammation and respiratory failure. The binding of the COVID-19 protein to ACE-2 has been shown to downregulate its enzymatic activity and decrease angiotensin production (28). This mechanism may be involved in the pathogenesis of pulmonary hypertension and insufficiency caused by SARS-CoV-2 infection (29).

The inflammatory process is accompanied by the activation of the coagulation system, which is characterized by the interaction of coagulation factors, and as observed in severe cases of sepsis, could lead to a disseminated intravascular coagulation process. As recent studies described, COVID-19 is commonly complicated with coagulopathy, and disseminated intravascular coagulation (DIC) may exist in the majority of deaths. In addition to pulmonary pneumonia and inflammation, the presence of vascular dysfunction and thrombosis have been noted in severe COVID-19 patients (30-32).

Because ACE-2, the receptor necessary for virus uptake, is expressed in the cardiovascular system (33), it is expected that this tissue is also susceptible to SARS-CoV-2 infection. Some new studies showed that COVID induces a vascular process in the lung, which includes vasodilation and endothelial dysfunction (34). In this regard, Caruso et al. (35) observed subsegmental vascular enlargement via computer tomography of patients with COVID-19. The presence of thrombosis or acute pulmonary embolism in COVID-19 patients associated with increased respiratory dead space was also reported by Chen et al. (36).

Similar to the disease caused by SARS-CoV identified in 2002, pulmonary involvement is the dominant clinical feature observed in COVID-19. Zhang et al. (30) observed the presence of Rp3 NP SARS-CoV-2 protein in alveolar epithelium cells, including those that were peeled and injured in the alveolar space in lung biopsies. These authors also observed diffuse alveolar damage, epithelial cells peeling with type II reactive hyperplasia of pneumocytes, and fibrous exudate associated with interstitial fibrosis and chronic inflammatory infiltrate.

Radiological findings in SARS-CoV-2-infected patients have been well-reported in several articles, and the consensus is that COVID-19 induces pneumonia with bilateral

lung involvement and ground-glass opacity, the latter being one of the most important radiological characteristic of COVID-19 that can be present even in asymptomatic patients (30,37-39).

According to Mason (40), the pathogenesis of COVID-19 can be divided into three stages. The first asymptomatic phase occurs when the inhaled virus SARS-CoV-2 starts to replicate in the epithelial cells in the nasal cavity. The second phase occurs when the virus migrates through the lung and a more vigorous innate immune response is triggered. The last phase is manifested by hypoxia, ground glass infiltrate, and progression to respiratory failure. A range of 20% to 30% of infected patients will progress to stage 3 of the disease and will develop pulmonary infiltrates that can also evolve into acute respiratory distress syndrome (ARDS). ARDS is an acute inflammatory lung injury, characterized by increased pulmonary vascular permeability, enlarged lung weight, and loss of aerated lung tissue (41). ARDS in its most severe form presents with intense pulmonary inflammation where there is severe hypoxemia (42). In terms of diagnosis, ARDS is characterized by a PaO₂/FiO₂ ratio equal to or less than 200 mmHg. In essence, the mechanisms underlying the triggering of ARDS are limited to damage to the alveolar epithelium and capillary endothelium (43,44). There is activation of neutrophils and high levels of proinflammatory cytokine release as well as activation of M1-like macrophages and reactive oxygen and nitrogen species. The incidence of ARDS is higher in older patients (45), and aging of the lungs may contribute to the higher mortality rate due to SARS-CoV-2 in elderly and immunosuppressed individuals.

The severe form of COVID-19 is well-characterized by dyspnea, hypoxemia, and pulmonary tissue damage. In addition, 67-85% of critically ill patients admitted to the intensive care unit (ICU) with COVID-19 will develop ARDS, and this will be the cause of death (46,47). Huang et al. (46) reported that dyspnea and lymphopenia was present in more than 50% of infected patients, and all of them showed abnormal findings on a chest CT scan. Considering the inflammatory responses, Lagunas-Rangel et al. (48) suggested that patients with severe COVID-19 show a cytokine storm with acute increased levels of IL-2, IL-7, G-CSF, CXCL10, MCP-1, MIP-1 α , TNF- α , and IL-6, similar to patients diagnosed with SARS and MERS. Thus, this suggests that mortality could be due to viral hyperinflammation that exacerbates lung damage.

Some evidence associated high IL-6 levels with the severity and mortality of COVID-19 (49,50), although the levels of interferon (IFN)- γ tend to be somewhat decreased in severe cases compared with moderate cases. These cytokines are also increased in both the serum and bronchoalveolar lavage as well as in lung tissue from ARDS patients (51-53). This alteration in inflammatory response observed in COVID-19 patients could be associated with a reduction in CD4⁺, CD8⁺, and NK lymphocytes (36). In this regard, CD4⁺ T and CD8⁺ T were strongly reduced in patients with severe COVID-19 compared with those in patients with mild disease (54).

Sex differences in immune response and inflammation

Steroid sex hormones may affect the strength of immune responses in opposite directions and result in a general difference between males and females, with stronger immune



responses in females than in males (for review see Roved et al. (10)). The dominant profile of the immune response may be dependent on hormonal variation during the female sexual cycle. Data indicate that female sex hormones enhance Th2 immune responses and reduce Th1, and this influence has been comprehensively covered in many reviews (9,55,56). Females seem to develop stronger cell-dependent and humoral responses to infection and vaccination. The result of these differences can be observed when comparing the incidence of autoimmune diseases between sexes (57), which is higher in females, and also by the observation of higher incidence of sepsis in males in parallel with higher plasma levels of Th1 cytokines (58-60). Also, the incidence of sepsis in postmenopausal women increases to levels almost equal to those seen in age-matched men (61). In addition to hormonal influences, there is an apparent genetic disparity, since females carry two inherently polymorphic X chromosomes, while males have only one polymorphic X chromosome passed from the mother. The random process of X-chromosome inactivation provides females with a broader panel of proteins and enlarges the potential diversity of cell populations (9).

In women, the CD4⁺ T cell numbers are higher (62), and the ovarian cycle also influences the T reg cell populations with an increased number when estradiol levels are higher (63). These differences are connected to estradiol influence and affect immune responses to infection. In addition, estradiol exerts anti-inflammatory effects on innate immune responses by reducing monocyte and macrophage inflammatory cytokine release (64), delaying neutrophil apoptosis and enhancing neutrophil annexin-1 expression without increasing their activation (65,66). Experimental data confirm that the leukocyte function in females is more efficient than that in males, detecting and eliminating pathogens more rapidly, and that females have higher numbers of tissue macrophages with a greater density of toll-like receptors; female macrophage phagocytosis is more efficient with NADPH oxidase killing, in parallel with an increased population of resident anti-inflammatory T-lymphocytes (67). The dendritic cell population is also influenced by female sex hormones, with estradiol signaling via estrogen receptor α ER- α to increase the number of new DCs during inflammation (68). In plasmacytoid dendritic cells, which are mostly involved in antiviral responses, estrogen increases the release of type 1 cytokines that are important for an efficient type 1 immune response (69). The action of estradiol via ER- α also promotes activation of type 1 INF-inducible innate pathways, and the IFN pathway genes are highly expressed in females, regulating innate immunity (55).

Considering lung inflammation, mortality in ARDS is higher in males compared to females (70,71). In the respiratory system, sex steroids have been demonstrated to play important roles in the development and maturation of lungs and maintenance of normal lung function (72), and sexual dimorphism impacts the prevalence and incidence of several lung diseases (73). Sex differences were identified in lung physiology and disease and have been studied in animal models. Sex also influences lung development and disease conditions (74). Sex hormones may contribute to the disease pathophysiology or serve as protective factors, depending on the disease involved. Women may be protected from certain age-related biological processes by producing lower levels of reactive oxidant species, important drivers of pathology in age-related pulmonary disease (75). Pulmonary diseases in

women are influenced by age because of the variation of their level of estrogen. Increasing age is associated with higher levels of circulating inflammatory mediators and acute phase proteins. The reduction of estrogen, the increased fat tissue, and the presence of subclinical infections contribute to the proinflammatory status of postmenopausal women. The development and progression of age-related diseases is dependent on mechanisms associated with cellular senescence that may be triggered by telomere shortening, which in women occurs at a slower rate than in men (76).

Substantial evidence supports the anti-inflammatory role of systemic estrogens. The mechanisms behind the female protection from infection are reported to be mediated mainly by sex hormones, in particular 17 β -estradiol, that can directly influence synthesis and signal transduction of multiple cytokines *in vitro* (67).

17 β -estradiol therapeutic effects in experimental models

Experimental investigations of several disease models confirm that higher levels of systemic estrogens promote anti-inflammatory responses. 17- β -estradiol is the predominant estrogen during the reproductive years, both in its total serum concentration and in overall estrogenic activity. Estradiol down-regulates immune functions in endothelial cells, including the stimulation of leukocyte adhesion and migration to infected tissues (77-79). It is known that estrogen plays a protective role in endothelial function (80), activating the generation of nitric oxide (NO) via endothelial nitric oxide synthase (eNOS), which is mediated by mechanisms that lead to an increase in NO bioavailability through the induction of gene transcription and activation of eNOS via phosphatidylinositol-3-kinase/serine-threonine kinase (PI3K/AKT) (81). Still, studies by our group using an aortic occlusion model in male rats indicate that previous treatment with estradiol has a beneficial effect on the course of mesenteric ischemia and intestinal injury, preventing mortality due to systemic inflammation (82). Additionally, in an intestinal ischemia and reperfusion (I/R) model, results indicate that female sex hormones, notably estradiol, exert a protective effect on preventing/reducing lung and intestinal injuries caused by systemic inflammation after ischemic trauma (83-85). Proestrus or estradiol-treated females showed reduced lung damage and inflammation in ischemia and reperfusion models (86,87) and in a hemorrhagic shock model (88). In addition, treatment with estradiol showed anti-inflammatory action similar to that of glucocorticoids, with reduced expression of transcription factors involved in the inflammatory response and reduced recruitment of neutrophils by decreasing the production of interleukins, such as IL-8, in addition to chemokines and adhesion molecules (66,89).

Evidence from clinical and experimental studies strongly suggests that estrogens can modulate lung inflammation and allergic reactions, because activation of estrogen receptors modulates immune cells and both innate and adaptive immune responses (55,73). However, the effects of estradiol as a pro or anti-inflammatory factor seem to be dependent on the pathogenesis of the diseases studied. Estradiol plays a critical role in improving outcomes in the settings of trauma, shock, sepsis, myocardial ischemia/reperfusion, and acute lung injury (ALI). Experimental data with an intestinal I/R model show that estradiol treatment in ovariectomized



females reduces lung inflammation and exerts these effects by modulating eNOS protein expression in the lungs (83). As permeability edema represents a life-threatening complication of ALI, estradiol effects on the control of lung vascular permeability could be considered among the therapeutic strategies to reduce lung edema (83). In this model, the inflammatory status of the lungs in ovariectomized females remains for a long period after the mesenteric reperfusion has been reestablished and is characterized by the release of proinflammatory mediators and danger signals. The treatment with estradiol is able to downregulate the lung inflammation and its capacity for releasing cytokines (90). In males, using the same I/R model, estradiol treatment was effective in reducing lung inflammation even at 1h after the reestablishment of intestinal perfusion (84). Thus, the effects of estradiol are important in the modulation of an already established lung inflammatory response. It is noteworthy to point out that the proposed treatment was acute and could be considered a clinical alternative.

Estrogen attenuates the vasoconstrictor response to various stimuli and induces vasodilation in the pulmonary vasculature during stress situations like hypoxia (91). This is mediated by increased levels of prostacyclin and NO as well as decreased levels of endothelin-1 (ET-1). It is well-established that estradiol exerts important vasoactive effects by activation of eNOS, as well as modulation of vasoactive substances released from endothelial cells or the direct vasodilation effect via relaxation of smooth muscle cells (92). The vasoactive effects of ET-1 can be influenced by estradiol and are organ-dependent (93), and estradiol and its metabolites inhibit ET-1 synthesis (94). Ovariectomized female rats present higher ET-1, and estradiol replacement reduces ET-1 peptide expression (95). Thus, estradiol acts as a protective vasoactive agent against deleterious microcirculatory conditions, such as brain death and ischemia/reperfusion injury (82,96). Brain-dead rats treated with 17 β -estradiol exhibit reduced lung injury, mainly because of its actions on endothelial and inducible nitric oxide synthase (iNOS) (96). Regarding injury, estrogen can inhibit vascular responses and thus prevent proatherosclerotic events (97). The maintenance of organ homeostasis depends on a complex network of systems, and reduction of blood flow may have profound consequences on organ status. Microcirculatory function following trauma can be affected by sex hormones (98), altering tissue perfusion and influencing inflammatory processes. Estrogen exerts a variety of rapid actions, initiated after coupling with membrane receptors, which may positively modulate vascular responses in pulmonary disease and help to maintain microvascular flow.

COVID-19, coagulation, and estradiol influence

Although one of the consequences of severe COVID-19 is the development of ARDS, some studies indicated that respiratory phenotypes induced by COVID-19 are slightly different from those classical phenotypes observed in ARDS patients (99). COVID-19 patients developed a severe hypoxemia early in the course of the disease, which is silent most of the time (100). Most recently, small fibrinous thrombi in small pulmonary arterioles and a tumefaction of endothelial were observed in autopsies in cases of fatal COVID-19 (101). This could indicate the activation of the coagulation cascade in these patients. In addition, they also reported diffuse alveolar damage with intense epithelial viral cytopathic

effects both in the alveolar and small airways. More interestingly, the presence of thrombus occurred in both damaged and preserved lung parenchyma areas. All together, these features suggested that anticoagulant drugs could have beneficial effects in COVID-19 patients.

Estrogen reduces platelet aggregation, the opposite effect of testosterone (102,103). Notelovitz et al. (104) studied the short-term effects of estrogens on the coagulation-fibrinolysis process in surgically treated women, and the results did not show abnormal or thrombogenic changes in coagulation parameters. *In vitro*, 17 β -estradiol is able to inhibit platelet aggregation by promoting Ca²⁺ extrusion or reuptake activity, and its action is dependent on the increase of NO synthesis (105). Platelet reactivity plays a pivotal role in thrombus formation, and *in vitro* data suggest that although women have a higher magnitude of platelet reactivity, their response to aspirin is similar or even larger compared to that of men (106). Experimental data show that a chronic high physiologic level of estrogen equivalent to that observed in pregnant mice had a significant inhibitory effect on platelet aggregation (107). Estrogen mediates beneficial effects on the cerebral microcirculation and moderated cerebral thrombotic mechanisms by enhancing the plasma levels of NO (108). Moreover, estrogens exert positive effects on the pulmonary vasculature by increasing prostacyclin release and NO production by eNOS (109). Cowman et al. (110) evidenced a connection between age and decreases in platelet tracks, platelet translocation, and unstable platelet interactions. The altered platelet function associated with aging was more profound in females compared with that of males and could be a result of a reduction in estrogen levels in women.

The role of estrogen and estrogen-related compounds in antiviral mechanism activity

Considering virus-induced lung inflammation, the excessive inflammation leads to tissue damage. In this regard, T cells seems to be reduced in COVID-19 patients, and the present T cells seemed to be functionally exhausted (111). Severe COVID-19 can induce ARDS in more than 50% of patients, and neutrophils are considered to be the central cells involved in the pathogenesis of ARDS. Infected female mice that were administered estradiol showed pulmonary recruitment of neutrophils and virus-specific CD8 T cells, releasing more INF- γ and TNF- α (112). Moreover, exogenous estrogen treatment in female mice infected with H1N1 reduced total pulmonary inflammation and the levels of pro-inflammatory genes in the lung (113).

Many viruses are affected by estrogen at the molecular level, specifically in the replication machinery and maturation of the virion, but to date, no evidence has been presented for SARS-CoV-2. Despite the well-known role of estrogen in inflammation to combat pathogens and infections, it has been shown that estrogen receptors participate in repressing transcription virus genes. One example is the hepatitis B virus (HBV) infection, where ER- α interferes with virus gene expression through (HNF)-4 α (hepatocyte nuclear factor 4 α) binding to HBV enhancer I in hepatocytes (114). In the hepatitis C virus (HCV) infection model *in vitro* (Huh7 cells), 17 β -estradiol reduced virus infection by interfering with assembly and/or virus release, disrupting the virus life cycle, and this effect was inhibited by anti-estrogen Fulvestrant (115). Moreover, HCV-infected women, after postmenopause, exhibit a reduced response to antiviral therapy and

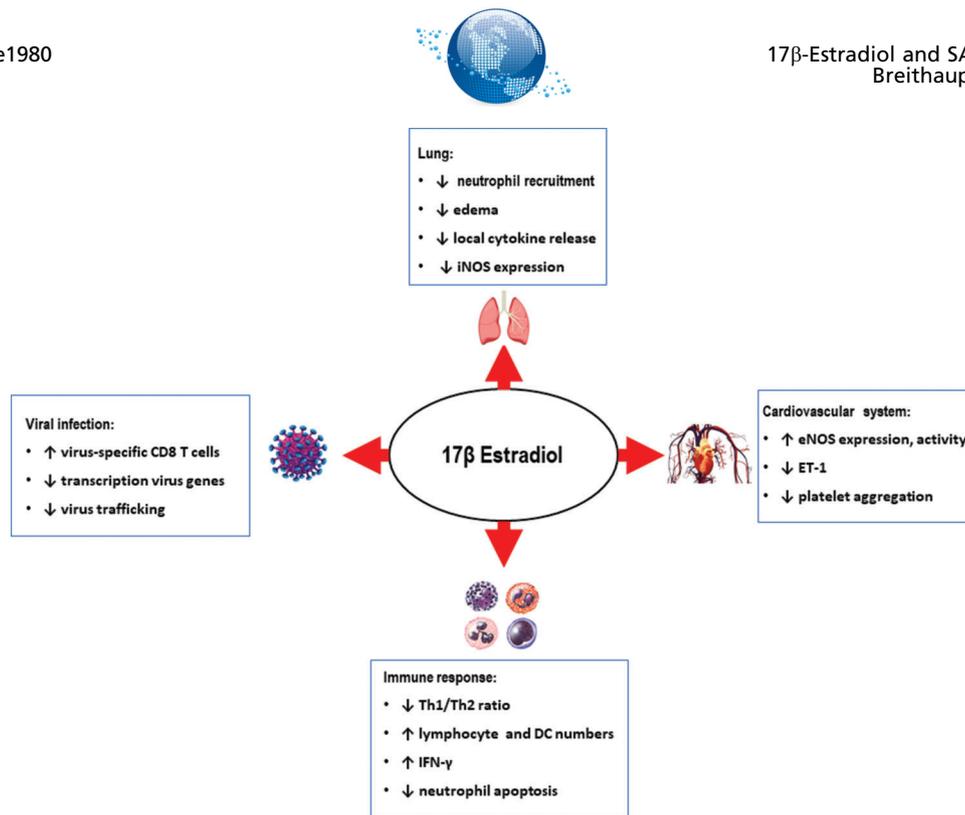


Figure 1 - Effects of 17 β -estradiol in different compartments/systems.

greater incidence of hepatic fibrosis and hepatic carcinoma (116,117).

Nevertheless, the most likely antiviral effects of estrogen are mediated by inflammation modulation. Regarding the cells that participate in the immune response, dendritic cells play a crucial role. Escribese et al. (118) infected dendritic cells with Newcastle disease virus (NDV) and treated the cells with 17 β -estradiol. The authors observed a diminished type I IFN synthesis, also reducing the CD4 T cell response, suggesting that estrogen has an antiviral activity on RNA viruses.

According to the evidence that female sex hormones exert antiviral activity in many organs, the use of selective estrogen receptor modulators (SERMs) has been extensively explored in experimental systems to reduce virus infections (reviewed by Montoya & Krysan, (119)). In general, SERMs have been used for several therapeutic purposes, such as in the treatment for infertility and osteoporosis, but are mainly related to cancer as adjuvants in chemotherapy, such as tamoxifen. In viral infection, this compound was suggested to be considered as an antiviral therapy, because of its properties of disrupting viral replication during HIV (human immunodeficiency virus) infection in monocytes and CD4 + T-lymphocyte cell lines (120). Tamoxifen has also displayed a capacity to inhibit HCV replication via estrogen receptor assembly to RNA-dependent RNA polymerase NS5B (121,122). Murakami et al. (122) showed that various SERMs reduce the production of HCV RNA, preferentially related to the extracellular but also intracellular virus proteins, suggesting that SERMs may be a multi-targeted drug that interferes with several processes, comprising virus entry and replication.

Of note, the most approximate model of SARS-CoV-2 viral infection is found in MERS-CoV and SARS-CoV. Both have been demonstrated to be affected by toremifene citrate (a classical SERM), which delayed virus infection due to the

inhibition of the late endosome virus trafficking (123), and a similar effect was observed previously with Ebola virus infection (124). In addition, both *in vitro* and *in vivo* experiments showed that toremifene and clomifene present antiviral activity against a large variety of Ebola virus strains (125,126). Importantly, it was also demonstrated that SERMs can elevate the survival rate of infected animals by 90% and 50% when using clomifene and toremifene, respectively. Mechanistically, it has been proposed that toremifene could target envelope proteins GP1 and GP2, which permit the viral attachment and access to host cells, decreasing its stability and resulting in reduced endolysosomal membrane fusion (127). This may be considered good evidence to support that estrogen and estrogen-related compounds play a major role in antiviral therapies for SARS-CoV-2.

■ CONCLUSION

Abundant literature highlights sex differences in immune response and its influence on the incidence and severity of diseases. Considering trauma, shock, and infection, the female sex is associated with advantageous outcomes. Since the hormonal context exerts an important influence on the homeostasis control and the defense mechanisms, it is important to consider the beneficial effects of estrogens with regard to providing better control of defense and the different immune cells involved, in addition to cardiovascular system protection and flow maintenance. Direct and indirect mechanisms underlying the effects of estradiol were investigated, and the results point to a possible positive effect of estradiol as an adjuvant therapeutic element for the treatment of patients affected by the novel coronavirus, SARS-CoV-2 (Figure 1). Studies are being carried out in order to analyze the effects estradiol on SARS-CoV-2 *in vitro*, and in parallel, there is already a clinical trial in course.



AUTHOR CONTRIBUTIONS

All of the authors conceived the study and were responsible for the manuscript drafting, editing, and review.

REFERENCES

- Sanders JM, Monogue ML, Jodlowski TZ, Cutrell JB. Pharmacologic Treatments for Coronavirus Disease 2019 (COVID-19): A Review. *JAMA*. 2020. <https://doi.org/10.1001/jama.2020.6019>
- Yang X, Yu Y, Xu J, Shu H, Xia J, Liu H, et al. Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered, retrospective, observational study. *Lancet Respir Med*. 2020;8(5):475-81. [https://doi.org/10.1016/S2213-2600\(20\)30079-5](https://doi.org/10.1016/S2213-2600(20)30079-5)
- Chen T, Wu D, Chen H, Yan W, Yang D, Chen G, et al. Clinical characteristics of 113 deceased patients with coronavirus disease 2019: retrospective study. *BMJ*. 2020;368:m1091. <https://doi.org/10.1136/bmj.m1091>
- Guan WJ, Ni ZY, Hu Y, Liang WH, Ou CQ, He JX, et al. Clinical Characteristics of Coronavirus Disease 2019 in China. *N Engl J Med*. 2020;382(18):1708-20. <https://doi.org/10.1056/NEJMoa2002032>
- Global Health [Internet]. Available from: <https://globalhealth5050.org/covid19/>
- La Vignera S, Cannarella R, Condorelli RA, Torre F, Aversa A, Calogero AE. Sex-Specific SARS-CoV-2 Mortality: Among Hormone-Modulated ACE2 Expression, Risk of Venous Thromboembolism and Hypovitaminosis D. *Int J Mol Sci*. 2020;21(8). pii: E2948. <https://doi.org/10.3390/ijms21082948>
- New York Health [Internet]. Available from: <https://www1.nyc.gov/site/doh/covid/covid-19-data.page>
- Chen H, Guo J, Wang C, Luo F, Yu X, Zhang W, et al. Clinical characteristics and intrauterine vertical transmission potential of COVID-19 infection in nine pregnant women: a retrospective review of medical records. *Lancet*. 2020;395(10226):809-15. [https://doi.org/10.1016/S0140-6736\(20\)30360-3](https://doi.org/10.1016/S0140-6736(20)30360-3)
- Fish EN. The X-files in immunity: sex-based differences predispose immune responses. *Nat Rev Immunol*. 2008;8(9):737-44. <https://doi.org/10.1038/nri2394>
- Roved J, Westerdaal H, Hasselquist D. Sex differences in immune responses: Hormonal effects, antagonistic selection, and evolutionary consequences. *Horm Behav*. 2017;88:95-105. <https://doi.org/10.1016/j.yhbeh.2016.11.017>
- Grossman CJ. Interactions between the gonadal steroids and the immune system. *Science*. 1985;227(4684):257-61. <https://doi.org/10.1126/science.3871252>
- Olsen NJ, Kovacs WJ. Gonadal steroids and immunity. *Endocr Rev*. 1996;17(4):369-84. <https://doi.org/10.1210/edrv-17-4-369>
- Schröder J, Kahlke V, Book M, Stüber F. Gender differences in sepsis: genetically determined? *Shock*. 2000;14(3):307-10. <https://doi.org/10.1097/00024382-200014030-00011>
- Al-Tarrah K, Moiemien N, Lord J. The influence of sex steroid hormones on the response to trauma and burn injury. *Burn Trauma*. 2017;5:29. <https://doi.org/10.1186/s41038-017-0093-9>
- Angele MK, Schwacha MG, Ayala A, Chaudry IH. Effect of gender and sex hormones on immune responses following shock. *Shock*. 2000;14(2):81-90. <https://doi.org/10.1097/00024382-200014020-00001>
- Angele MK, Frantz MC, Chaudry IH. Gender and sex hormones influence the response to trauma and sepsis: potential therapeutic approaches. *Clinics*. 2006;61(5):479-88. <https://doi.org/10.1590/S1807-59322006000500017>
- Chaudry IH, Samy TS, Schwacha MG, Wang P, Rue LW, Bland KI. Endocrine targets in experimental shock. *J Trauma*. 2003;54(5 Suppl):S118-25. <https://doi.org/10.1097/01.TA.0000064511.14322.F1>
- Frink M, Pape HC, van Griensven M, Krettek C, Chaudry IH, Hildebrand F. Influence of sex and age on mods and cytokines after multiple injuries. *Shock*. 2007;27(2):151-6. <https://doi.org/10.1097/01.shk.0000239767.64786.de>
- Hildebrand F, Thobe BM, Hubbard WJ, Choudhry MA, Pape HC, Chaudry IH. Effects of 17beta-estradiol and flutamide on splenic macrophages and splenocytes after trauma-hemorrhage. *Cytokine*. 2006;36(3-4):107-14. <https://doi.org/10.1016/j.cyto.2006.11.002>
- Hsieh YC, Frink M, Thobe BM, Hsu JT, Choudhry MA, Schwacha MG, et al. 17Beta-estradiol downregulates Kupffer cell TLR4-dependent p38 MAPK pathway and normalizes inflammatory cytokine production following trauma-hemorrhage. *Mol Immunol*. 2007;44(9):2165-72. <https://doi.org/10.1016/j.molimm.2006.11.019>
- Knöferl MW, Jarrar D, Angele MK, Ayala A, Schwacha MG, Bland KI, et al. 17 beta-Estradiol normalizes immune responses in ovariectomized females after trauma-hemorrhage. *Am J Physiol Cell Physiol*. 2001;281(4):C1131-8. <https://doi.org/10.1152/ajpcell.2001.281.4.C1131>
- Knöferl MW, Angele MK, Diodato MD, Schwacha MG, Ayala A, Cioffi WG, et al. Female sex hormones regulate macrophage function after trauma-hemorrhage and prevent increased death rate from subsequent sepsis. *Ann Surg*. 2002;235(1):105-12. <https://doi.org/10.1097/0000658-200201000-00014>
- Moeinpour F, Choudhry MA, Kawasaki T, Timares L, Schwacha MG, Bland KI, et al. 17 Beta-estradiol normalizes Toll receptor 4, mitogen activated protein kinases and inflammatory response in epidermal keratinocytes following trauma-hemorrhage. *Mol Immunol*. 2007;44(13):3317-23. <https://doi.org/10.1016/j.molimm.2007.02.023>
- Stangl V, Baumann G, Stangl K. Coronary atherogenic risk factors in women. *Eur Heart J*. 2002;23(22):1738-52. <https://doi.org/10.1053/eurh.2002.3329>
- Suba Z. Prevention and therapy of COVID-19 via exogenous estrogen treatment for both male and female patients. *J Pharm Pharm Sci*. 2020;23(1):75-85. <https://doi.org/10.18433/jpps31069>
- Xu X, Chen P, Wang J, Feng J, Zhou H, Li X, et al. Evolution of the novel coronavirus from the ongoing Wuhan outbreak and modeling of its spike protein for risk of human transmission. *Sci China Life Sci*. 2020;63(3):457-60. <https://doi.org/10.1007/s11427-020-1637-5>
- Wujtewicz M, Dylczyk-Sommer A, Aszkielowicz A, Zdanowski S, Piwowarczyk S, Owczuk R. COVID-19 - what should anaesthesiologists and intensivists know about it? *Anaesthesiol Intensive Ther*. 2020;52(1):34-41. <https://doi.org/10.5114/ait.2020.93756>
- Mo P, Xing Y, Xiao Y, Deng L, Zhao Q, Wang H, et al. Clinical characteristics of refractory COVID-19 pneumonia in Wuhan, China. *Clin Infect Dis*. 2020. pii: ciaa270. <https://doi.org/10.1093/cid/ciaa270>
- Gurwitz D. Angiotensin receptor blockers as tentative SARS-CoV-2 therapeutics. *Drug Dev Res*. 2020. <https://doi.org/10.1002/ddr.21656>
- Zhang H, Zhou P, Wei Y, Yue H, Wang Y, Hu M, et al. Histopathologic Changes and SARS-CoV-2 Immunostaining in the Lung of a Patient With COVID-19. *Ann Intern Med*. 2020. <https://doi.org/10.7326/M20-0533>
- Barrett CD, Moore HB, Yaffe MB, Moore EE. ISTH interim guidance on recognition and management of coagulopathy in COVID-19: A Comment. *J Thromb Haemost*. 2020. <https://doi.org/10.1111/jth.14860>
- Connors JM, Levy JH. COVID-19 and its implications for thrombosis and anticoagulation. *Blood*. 2020. pii: blood.202006000. <https://doi.org/10.1182/blood.202006000>
- Gheblawi M, Wang K, Viveiros A, Nguyen Q, Zhong JC, Turner AJ, et al. Angiotensin Converting Enzyme 2: SARS-CoV-2 Receptor and Regulator of the Renin-Angiotensin System: Celebrating the 20th Anniversary of the Discovery of ACE2. *Circ Res*. 2020;126(10):1456-74. <https://doi.org/10.1161/CIRCRESAHA.120.317015>
- Leisman DE, Deutschman CS, Legrand M. Facing COVID-19 in the ICU: vascular dysfunction, thrombosis, and dysregulated inflammation. *Intensive Care Med*. 2020. <https://doi.org/10.1007/s00134-020-06059-6>
- Caruso D, Zerunian M, Polici M, Pucciarelli F, Polidori T, Rucci C, et al. Chest CT Features of COVID-19 in Rome, Italy. *Radiology*. 2020. <https://doi.org/10.1148/radiol.2020201237>
- Chen G, Wu D, Guo W, Cao Y, Huang D, Wang H, et al. Clinical and immunological features of severe and moderate coronavirus disease 2019. *J Clin Invest*. 2020;30(5):2620-9. <https://doi.org/10.1172/JCI137244>
- Shi H, Han X, Jiang N, Cao Y, Alwalid O, Gu J, et al. Radiological findings from 81 patients with COVID-19 pneumonia in Wuhan, China: a descriptive study. *Lancet Infect Dis*. 2020;20(4):425-34. [https://doi.org/10.1016/S1473-3099\(20\)30086-4](https://doi.org/10.1016/S1473-3099(20)30086-4)
- Lai CC, Shih TP, Ko WC, Tang HJ, Hsueh PR. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and coronavirus disease-2019 (COVID-19): The epidemic and the challenges. *Int J Antimicrob Agents*. 2020;55(3):105924. <https://doi.org/10.1016/j.ijantimicag.2020.105924>
- Xu X, Yu C, Qu J, Zhang L, Jiang S, Huang D, et al. Imaging and clinical features of patients with 2019 novel coronavirus SARS-CoV-2. *Eur J Nucl Med Mol Imaging*. 2020;47(5):1275-80. <https://doi.org/10.1007/s00259-020-04735-9>
- Mason RJ. Pathogenesis of COVID-19 from a cell biologic perspective. *Eur Respir J*. 2020;55(4) pii: 2000607. <https://doi.org/10.1183/13993003.00607-2020>
- ARDS Definition Task Force, Ranieri VM, Rubenfeld GD, Thompson BT, Ferguson ND, Caldwell E, et al. Acute respiratory distress syndrome: the Berlin Definition. *JAMA*. 2012;307(23):2526-33. <https://doi.org/10.1001/jama.2012.5669>
- Bernard GR, Artigas A, Brigham KL, Carlet J, Falke K, Hudson L, et al. The American-European Consensus Conference on ARDS. Definitions, mechanisms, relevant outcomes, and clinical trial coordination. *Am J Respir Crit Care Med*. 1994;149(3 Pt 1):818-24. <https://doi.org/10.1164/ajrccm.149.3.7509706>
- Randhawa R, Bellingan G. Acute lung injury. *Anaesth Intensive Care Med*. 2007;8(11):477-80. <https://doi.org/10.1016/j.mpaic.2007.09.003>
- Matthay MA, Zemans RL, Zimmerman GA, Arabi YM, Beitler JR, Mercat A, et al. Acute respiratory distress syndrome. *Nat Rev Dis Prim*. 2019;5(1):18.
- Hecker L. Mechanisms and consequences of oxidative stress in lung disease: therapeutic implications for an aging populace. *Am J Physiol*



- Lung Cell Mol Physiol. 2018;314(4):L642-L653. <https://doi.org/10.1152/ajplung.00275.2017>
46. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020;395(10223):497-506. [https://doi.org/10.1016/S0140-6736\(20\)30183-5](https://doi.org/10.1016/S0140-6736(20)30183-5)
 47. Ruan Q, Yang K, Wang W, Jiang L, Song J. Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China. *Intensive Care Med*. 2020. <https://doi.org/10.1007/s00134-020-05991-x>
 48. Lagunas-Rangel FA, Chávez-Valencia V. High IL-6/IFN- γ ratio could be associated with severe disease in COVID-19 patients. *J Med Virol*. 2020. <https://doi.org/10.1002/jmv.25900>
 49. Zhou Y, Han T, Chen J, Hou C, Hua L, He S, et al. Clinical and Auto-immune Characteristics of Severe and Critical Cases with COVID-19. *Clin Transl Sci*. 2020. <https://doi.org/10.1111/cts.12805>
 50. Wu C, Chen X, Cai Y, Xia J, Zhou X, Xu S, et al. Risk Factors Associated with Acute Respiratory Distress Syndrome and Death in Patients with Coronavirus Disease 2019 Pneumonia in Wuhan, China. *JAMA Intern Med*. 2020. <https://doi.org/10.1001/jamainternmed.2020.0994>
 51. Pires-Neto RC, Morales MM, Lencas T, Inforsato N, Duarte MIS, Amato MB, et al. Expression of acute-phase cytokines, surfactant proteins, and epithelial apoptosis in small airways of human acute respiratory distress syndrome. *J Crit Care*. 2013;28(1):111.e9-111.e15. <https://doi.org/10.1016/j.jcrc.2012.05.013>
 52. Parsons EC, Hough CL, Seymour CW, Cooke CR, Rubenfeld GD, Watkins TR, et al. Red blood cell transfusion and outcomes in patients with acute lung injury, sepsis and shock. *Crit Care*. 2011;15(5):R221. <https://doi.org/10.1186/cc10458>
 53. Schütte H, Lohmeyer J, Rosseau S, Ziegler S, Siebert C, Kielisch H, et al. Bronchoalveolar and systemic cytokine profiles in patients with ARDS, severe pneumonia and cardiogenic pulmonary oedema. *Eur Respir J*. 1996;9(9):1858-67. <https://doi.org/10.1183/09031936.96.09091858>
 54. Wan S, Yi Q, Fan S, Lv J, Zhang X, Guo L, et al. Relationships among lymphocyte subsets, cytokines, and the pulmonary inflammation index in coronavirus (COVID-19) infected patients. *Br J Haematol*. 2020;189(3):428-37. <https://doi.org/10.1111/bjh.16659>
 55. Kovats S. Estrogen receptors regulate innate immune cells and signaling pathways. *Cell Immunol*. 2015;294(2):63-9. <https://doi.org/10.1016/j.cellimm.2015.01.018>
 56. Khan D, Ansar Ahmed S. The Immune System Is a Natural Target for Estrogen Action: Opposing Effects of Estrogen in Two Prototypical Autoimmune Diseases. *Front Immunol*. 2016;6:635. <https://doi.org/10.3389/fimmu.2015.00635>
 57. Fairweather D, Frisancho-Kiss S, Rose NR. Sex differences in autoimmune disease from a pathological perspective. *Am J Pathol*. 2008;173(3):600-9. <https://doi.org/10.2353/ajpath.2008.071008>
 58. Offner PJ, Moore EE, Biffi WL. Male gender is a risk factor for major infections after Vsurgery. *Arch Surg*. 1999;134(9):935-8. <https://doi.org/10.1001/archsurg.134.9.935>
 59. Wichmann MW, Inthorn D, Andress HJ, Schildberg FW. Incidence and mortality of severe sepsis in surgical intensive care patients: the influence of patient gender on disease process and outcome. *Intensive Care Med*. 2000;26(2):167-72. <https://doi.org/10.1007/s001340050041>
 60. George RL, McGwin G Jr, Windham ST, Melton SM, Metzger J, Chaudry IH, et al. Age-related gender differential in outcome after blunt or penetrating trauma. *Shock*. 2003;19(1):28-32. <https://doi.org/10.1097/00024382-200301000-00006>
 61. Martin GS, Mannino DM, Eaton S, Moss M. The epidemiology of sepsis in the United States from 1979 through 2000. *N Engl J Med*. 2003;348(16):1546-54. <https://doi.org/10.1056/NEJMoa022139>
 62. Amadori A, Zamarchi R, De Silvestro G, Forza G, Cavatton G, Danieli GA, et al. Genetic control of the CD4/CD8 T-cell ratio in humans. *Nat Med*. 1995;1(12):1279-83. <https://doi.org/10.1038/nm1295-1279>
 63. Arruvito L, Sanz M, Banham AH, Fainboim L. Expansion of CD4+ CD25+ and FOXP3+ regulatory T cells during the follicular phase of the menstrual cycle: implications for human reproduction. *J Immunol*. 2007;178(4):2572-8. <https://doi.org/10.4049/jimmunol.178.4.2572>
 64. Kramer PR, Kramer SE, Guan G. 17 beta-estradiol regulates cytokine release through modulation of CD16 expression in monocytes and monocyte-derived macrophages. *Arthritis Rheum*. 2004;50(6):1967-75. <https://doi.org/10.1002/art.20309>
 65. Molloy EJ, O'Neill AJ, Grantham JJ, Sheridan-Pereira M, Fitzpatrick JM, Webb DW, et al. Sex-specific alterations in neutrophil apoptosis: The role of estradiol and progesterone. *Blood*. 2003;102(7):2653-9. <https://doi.org/10.1182/blood-2003-02-0649>
 66. Nadkarni S, Cooper D, Brancialeone V, Bena S, Perretti M. Activation of the annexin A1 pathway underlies the protective effects exerted by estrogen in polymorphonuclear leukocytes. *Arterioscler Thromb Vasc Biol*. 2011;31(11):2749-59. <https://doi.org/10.1161/ATVBAHA.111.235176>
 67. Scotland RS, Stables MJ, Madalli S, Watson P, Gilroy DW. Sex differences in resident immune cell phenotype underlie more efficient acute inflammatory responses in female mice. *Blood*. 2011;118(22):5918-27. <https://doi.org/10.1182/blood-2011-03-340281>
 68. Kovats S. Estrogen receptors regulate an inflammatory pathway of dendritic cell differentiation: mechanisms and implications for immunity. *Horm Behav*. 2012;62(3):254-62. <https://doi.org/10.1016/j.yhbeh.2012.04.011>
 69. Murphy K, Weaver C. *Janeway's Immunobiology*. 9th ed. Taylor & Francis Group L, editor. New York: Garland Science; 2017.
 70. Agarwal R, Aggarwal AN, Gupta D, Behera D, Jindal SK. Etiology and outcomes of pulmonary and extrapulmonary acute lung injury/ARDS in a respiratory ICU in north India. *Chest*. 2006;130(3):724-9. <https://doi.org/10.1378/chest.130.3.724>
 71. Moss M, Mannino DM. Race and gender differences in acute respiratory distress syndrome deaths in the United States: an analysis of multiple-cause mortality data (1979-1996). *Crit Care Med*. 2002;30(8):1679-85. <https://doi.org/10.1097/00003246-200208000-00001>
 72. Asavasupreechar T, Saito R, Miki Y, Edwards DP, Boonyaratankornkit V, Sasano H. Systemic distribution of progesterone receptor subtypes in human tissues. *J Steroid Biochem Mol Biol*. 2020;199:105599. <https://doi.org/10.1016/j.jsbmb.2020.105599>
 73. Fuentes N, Silveyra P. Endocrine regulation of lung disease and inflammation. *Exp Biol Med*. 2018;243(17-18):1313-22. <https://doi.org/10.1177/1535370218816653>
 74. Carey MA, Card JW, Voltz JW, Germolec DR, Korach KS, Zeldin DC. The impact of sex and sex hormones on lung physiology and disease: lessons from animal studies. *Am J Physiol Lung Cell Mol Physiol*. 2007;293(2):L272-8. <https://doi.org/10.1152/ajplung.00174.2007>
 75. Nawrot TS, Staessen JA, Gardner JP, Aviv A. Telomere length and possible link to X chromosome. *Lancet*. 2004;363(9408):507-10. [https://doi.org/10.1016/S0140-6736\(04\)15535-9](https://doi.org/10.1016/S0140-6736(04)15535-9)
 76. Gardner M, Bann D, Wyley L, Cooper R, Hardy R, Nitsch D, et al. Gender and telomere length: systematic review and meta-analysis. *Exp Gerontol*. 2014;51:15-27. <https://doi.org/10.1016/j.exger.2013.12.004>
 77. Alvarez A, Hermenegildo C, Issekutz AC, Esplugues JV, Sanz MJ. Estrogens inhibit angiotensin II-induced leukocyte-endothelial cell interactions in vivo via rapid endothelial nitric oxide synthase and cyclooxygenase activation. *Circ Res*. 2002;91(12):1142-50. <https://doi.org/10.1161/01.RES.0000046018.23605.3E>
 78. Gao H, Liang M, Bergdahl A, Hamrén A, Lindholm MW, Dahlman-Wright K, et al. Estrogen attenuates vascular expression of inflammation associated genes and adhesion of monocytes to endothelial cells. *Inflamm Res*. 2006;55(8):349-53. <https://doi.org/10.1007/s00011-006-5194-z>
 79. Geraldes P, Gagnon S, Hadjadj S, Merhi Y, Sirois MG, Cloutier J, et al. Estradiol blocks the induction of CD40 and CD40L expression on endothelial cells and prevents neutrophil adhesion: An ERalpha-mediated pathway. *Cardiovasc Res*. 2006;71(3):566-73. <https://doi.org/10.1016/j.cardiores.2006.05.015>
 80. Ma XL, Gao F, Chen J, Christopher TA, Lopez BL, Ohlstein EH, et al. Endothelial protective and antishock effects of a selective estrogen receptor modulator in rats. *Am J Physiol Heart Circ Physiol*. 2001;280(2):H876-84. <https://doi.org/10.1152/ajpheart.2001.280.2.H876>
 81. Novella S, Dantas AP, Segarra G, Medina P, Hermenegildo C. Vascular Aging in Women: is Estrogen the Fountain of Youth? *Front Physiol*. 2012;3:165. <https://doi.org/10.3389/fphys.2012.00165>
 82. Rocha de Sousa PT, Breithaupt-Faloppa AC, de Jesus Correia C, Simão RR, Ferreira SG, Fiorelli AI, et al. 17 β -Estradiol prevents mesenteric injury induced by occlusion of the proximal descending aorta in male rats. *J Vasc Surg*. 2018;67(2):597-606. <https://doi.org/10.1016/j.jvs.2016.12.125>
 83. Breithaupt-Faloppa AC, Fantozzi ET, de Assis Ramos MM, Vitoretto LB, Couto GK, Lino-dos-Santos-Franco A, et al. Protective effect of estradiol on acute lung inflammation induced by an intestinal ischemic insult is dependent on nitric oxide. *Shock*. 2013;40(3):203-9. <https://doi.org/10.1097/SHK.0b013e3182a01e24>
 84. Breithaupt-Faloppa AC, Thais Fantozzi E, Romero DC, Rodrigues Ada S, de Sousa PT, Lino Dos Santos Franco A, et al. Acute effects of estradiol on lung inflammation due to intestinal ischemic insult in male rats. *Shock*. 2014;41(3):208-13. <https://doi.org/10.1097/SHK.0000000000000092>
 85. Ricardo-Da-Silva FY, Fantozzi ET, Rodrigues-Garbin S, Oliveira-Filho RM, Vargaftig BB, Breithaupt-Faloppa AC, et al. Estradiol Modulates Local Gut Injury Induced by Intestinal Ischemia-Reperfusion in Male Rats. *Shock*. 2017;48(4):477-83. <https://doi.org/10.1097/SHK.00000000000000873>
 86. Deitch EA, Feketeova E, Lu Q, Zaets S, Berezina TL, Machiedo GW, et al. Resistance of the female, as opposed to the male, intestine to I/R-mediated injury is associated with increased resistance to gut-induced distant organ injury. *Shock*. 2008;29(1):78-83. <https://doi.org/10.1097/shk.0b013e318063e98a>
 87. Mrazkova H, Lischke R, Hergert J. Influence of gender on ischemia-reperfusion injury in lungs in an animal model. *Physiol Res*. 2016;65(6):953-8. <https://doi.org/10.33549/physiolres.933273>
 88. Raju R, Chaudry IH. Sex steroids receptor antagonist: their use as adjuncts after trauma-hemorrhage for improving immune/cardiovascular responses and for decreasing mortality from subsequent sepsis.



- Anesth Analg. 2008;107(1):159-66. <https://doi.org/10.1213/ane.0b013e318163213d>
89. Nadkarni S, McArthur S. Oestrogen and immunomodulation: new mechanisms that impact on peripheral and central immunity. *Curr Opin Pharmacol*. 2013;13(4):576-81. <https://doi.org/10.1016/j.coph.2013.05.007>
90. Fantozzi ET, Breithaupt-Faloppa AC, Ricardo-da-Silva FY, Rodrigues-Garbin S, Romero DC, da Silva Rodrigues A, et al. Estradiol mediates the long-lasting lung inflammation induced by intestinal ischemia and reperfusion. *J Surg Res*. 2018;221:1-7. <https://doi.org/10.1016/j.jss.2017.07.038>
91. Lahm T, Crisostomo PR, Markel TA, Wang M, Weil BR, Novotny NM, et al. The effects of estrogen on pulmonary artery vasoactivity and hypoxic pulmonary vasoconstriction: potential new clinical implications for an old hormone. *Crit Care Med*. 2008;36(7):2174-83. <https://doi.org/10.1097/CCM.0b013e31817d1a92>
92. Kuebler JF, Jarrar D, Toth B, Bland KI, Rue L 3rd, Wang P, et al. Estradiol administration improves splanchnic perfusion following trauma-hemorrhage and sepsis. *Arch Surg*. 2002;137(1):74-9. <https://doi.org/10.1001/archsurg.137.1.74>
93. Ba ZF, Lu A, Shimizu T, Szalay L, Schwacha MG, Rue W 3rd, et al. 17 β -Estradiol modulates vasoconstriction induced by endothelin-1 following trauma-hemorrhage. *Am J Physiol Heart Circ Physiol*. 2007;292(1):H245-50. <https://doi.org/10.1152/ajpheart.00809.2006>
94. Dubey RK, Jackson EK, Keller PJ, Imthurn B, Rosselli M. Estradiol metabolites inhibit endothelin synthesis by an estrogen receptor-independent mechanism. *Hypertension*. 2001;37(2 Pt 2):640-4. <https://doi.org/10.1161/01.HYP.37.2.640>
95. Akishita M, Ouchi Y, Miyoshi H, Orimo A, Kozaki K, Eto M, et al. Estrogen inhibits endothelin-1 production and c-fos gene expression in rat aorta. *Atherosclerosis*. 1996;125(1):27-38. [https://doi.org/10.1016/0021-9150\(96\)05836-4](https://doi.org/10.1016/0021-9150(96)05836-4)
96. Vieira RF, Breithaupt-Faloppa AC, Matsubara BC, Rodrigues G, Sanches MP, Armstrong-Jr R, et al. 17 β -Estradiol protects against lung injuries after brain death in male rats. *J Hear Lung Transplant*. 2018;37(11):1381-7. <https://doi.org/10.1016/j.healun.2018.06.015>
97. Arnal JF, Fontaine C, Billon-Galés A, Favre J, Laurell H, Lenfant F, et al. Estrogen receptors and endothelium. *Arterioscler Thromb Vasc Biol*. 2010;30(8):1506-12. <https://doi.org/10.1161/ATVBAHA.109.191221>
98. Sharawy N, Ribback S, Al-Banna N, Lehmann C, Kern H, Wendt M, et al. Estradiol receptors agonists induced effects in rat intestinal microcirculation during sepsis. *Microvasc Res*. 2013;85(1):118-27. <https://doi.org/10.1016/j.mvr.2012.10.002>
99. Cattinoni L, Chiumello D, Caironi P, Busana M, Romitti F, Brazzi L, et al. COVID-19 pneumonia: different respiratory treatments for different phenotypes? *Intensive Care Med*. 2020. <https://doi.org/10.1007/s00134-020-06033-2>
100. Xie J, Tong Z, Guan X, Du B, Qiu H, Slutsky AS. Critical care crisis and some recommendations during the COVID-19 epidemic in China. *Intensive Care Med*. 2020. <https://doi.org/10.1007/s00134-020-05979-7>
101. Dolhnikoff M, Duarte-Neto AN, de Almeida Monteiro RA, Ferraz da Silva LF, Pierre de Oliveira E, Nascimento Saldiva PH, et al. Pathological evidence of pulmonary thrombotic phenomena in severe COVID-19. *J Thromb Haemost*. 2020. <https://doi.org/10.1111/jth.14844>
102. Maccarrone M, Bari M, Battista N, Finazzi-Agrò A. Estrogen stimulates arachidonylethanolamide release from human endothelial cells and platelet activation. *Blood*. 2002;100(12):4040-8. <https://doi.org/10.1182/blood-2002-05-1444>
103. Selles J, Polini N, Alvarez C, Massheimer V. Progesterone and 17 β -estradiol acutely stimulate nitric oxide synthase activity in rat aorta and inhibit platelet aggregation. *Life Sci*. 2001;69(7):815-27. [https://doi.org/10.1016/S0024-3205\(01\)01174-2](https://doi.org/10.1016/S0024-3205(01)01174-2)
104. Notelovitz M, Kitchens CS, Ware MD. Coagulation and fibrinolysis in estrogen-treated surgically menopausal women. *Obstet Gynecol*. 1984;63(5):621-5.
105. Nakano Y, Oshima T, Matsuura H, Kajiyama G, Kambe M. Effect of 17 β -estradiol on inhibition of platelet aggregation in vitro is mediated by an increase in NO synthesis. *Arterioscler Thromb Vasc Biol*. 1998;18(6):961-7. <https://doi.org/10.1161/01.ATV.18.6.961>
106. Becker DM, Segal J, Vaidya D, Yanek LR, Herrera-Galeano JE, Bray PF, et al. Sex differences in platelet reactivity and response to low-dose aspirin therapy. *JAMA*. 2006;295(12):1420-7. <https://doi.org/10.1001/jama.295.12.1420>
107. Valéra MC, Gratacap MP, Gourdy P, Lenfant F, Cabou C, Toutain CE, et al. Chronic estradiol treatment reduces platelet responses and protects mice from thromboembolism through the hematopoietic estrogen receptor α . *Blood*. 2012;120(8):1703-12. <https://doi.org/10.1182/blood-2012-01-405498>
108. Ono H, Sasaki Y, Bamba E, Seki J, Giddings JC, Yamamoto J. Cerebral thrombosis and microcirculation of the rat during the oestrous cycle and after ovariectomy. *Clin Exp Pharmacol Physiol*. 2002;29(1-2):73-8. <https://doi.org/10.1046/j.1440-1681.2002.03600.x>
109. Morrell ED, Tsai BM, Crisostomo PR, Hammoud ZT, Meldrum DR. Experimental therapies for hypoxia-induced pulmonary hypertension during acute lung injury. *Shock*. 2006;25(3):214-26. <https://doi.org/10.1097/01.shk.0000191380.44972.46>
110. Cowman J, Dunne E, Oglesby I, Byrne B, Ralph A, Voisin B, et al. Age-related changes in platelet function are more profound in women than in men. *Sci Rep*. 2015;5:12235. <https://doi.org/10.1038/srep12235>
111. Chiappelli F, Khakshooy A, Greenberg G. CoViD-19 Immunopathology and Immunotherapy. *Bioinformation*. 2020;16(3):219-22. <https://doi.org/10.6026/97320630016219>
112. Robinson DP, Hall OJ, Nilles TL, Bream JH, Klein SL. 17 β -estradiol protects females against influenza by recruiting neutrophils and increasing virus-specific CD8 T cell responses in the lungs. *J Virol*. 2014;88(9):4711-20. <https://doi.org/10.1128/JVI.02081-13>
113. Vermillion MS, Ursin RL, Attreed SE, Klein SL. Estriol Reduces Pulmonary Immune Cell Recruitment and Inflammation to Protect Female Mice From Severe Influenza. *Endocrinology*. 2018;159(9):3306-20. <https://doi.org/10.1210/en.2018-00486>
114. Wang SH, Yeh SH, Lin WH, Yeh KH, Yuan Q, Xia NS, et al. Estrogen receptor α represses transcription of HBV genes via interaction with hepatocyte nuclear factor 4 α . *Gastroenterology*. 2012;142(4):989-998.e4. <https://doi.org/10.1053/j.gastro.2011.12.045>
115. Magri A, Barbaglia MN, Foglia CZ, Boccato E, Burlone ME, Cole S, et al. 17 β -estradiol inhibits hepatitis C virus mainly by interference with the release phase of its life cycle. *Liver Int*. 2017;37(5):669-77. <https://doi.org/10.1111/liv.13303>
116. Villa E, Karampatou A, Cammà C, Di Leo A, Luongo M, Ferrari A, et al. Early menopause is associated with lack of response to antiviral therapy in women with chronic hepatitis C. *Gastroenterology*. 2011;140(3):818-29. <https://doi.org/10.1053/j.gastro.2010.12.027>
117. Yu JW, Sun LJ, Zhao YH, Kang P, Yan BZ. Impact of sex on virologic response rates in genotype 1 chronic hepatitis C patients with peginterferon alpha-2a and ribavirin treatment. *Int J Infect Dis*. 2011;15(11):e740-6. <https://doi.org/10.1016/j.ijid.2011.05.018>
118. Escribese MM, Kraus T, Rhee E, Fernandez-Sesma A, López CB, Moran TM. Estrogen inhibits dendritic cell maturation to RNA viruses. *Blood*. 2008;112(12):4574-84. <https://doi.org/10.1182/blood-2008-04-148692>
119. Montoya MC, Krysan DJ. Repurposing Estrogen Receptor Antagonists for the Treatment of Infectious Disease. *mBio*. 2018;9(6). pii: e02272-18. <https://doi.org/10.1128/mBio.02272-18>
120. Laurence J, Cooke H, Sikder SK. Effect of tamoxifen on regulation of viral replication and human immunodeficiency virus (HIV) long terminal repeat-directed transcription in cells chronically infected with HIV-1. *Blood*. 1990;75(3):696-703. <https://doi.org/10.1182/blood.V75.3.696.696>
121. Watashi K, Inoue D, Hijikata M, Goto K, Aly HH, Shimotohno K. Anti-hepatitis C virus activity of tamoxifen reveals the functional association of estrogen receptor with viral RNA polymerase NS5B. *J Biol Chem*. 2007;282(45):32765-72. <https://doi.org/10.1074/jbc.M704418200>
122. Murakami Y, Fukasawa M, Kaneko Y, Suzuki T, Wakita T, Fukazawa H. Selective estrogen receptor modulators inhibit hepatitis C virus infection at multiple steps of the virus life cycle. *Microbes Infect*. 2013;15(1):45-55. <https://doi.org/10.1016/j.micinf.2012.10.003>
123. Dyal J, Coleman CM, Hart BJ, Venkataraman T, Holbrook MR, Kindrachuk J, et al. Repurposing of clinically developed drugs for treatment of Middle East respiratory syndrome coronavirus infection. *Antimicrob Agents Chemother*. 2014;58(8):4885-93. <https://doi.org/10.1128/AAC.03036-14>
124. Johansen LM, Brannan JM, Delos SE, Shoemaker CJ, Stossel A, Lear C, et al. FDA-approved selective estrogen receptor modulators inhibit Ebola virus infection. *Sci Transl Med*. 2013;5(190):190ra79. <https://doi.org/10.1126/scitranslmed.3005471>
125. Nelson EA, Barnes AB, Wiehle RD, Fontenot GK, Hoenen T, White JM. Clomiphene and Its Isomers Block Ebola Virus Particle Entry and Infection with Similar potency: Potential Therapeutic Implications. *Viruses*. 2016;8(8). pii: E206. <https://doi.org/10.3390/v8080206>
126. Madrid PB, Panchal RG, Warren TK, Shurtleff AC, Endsley AN, Green CE, et al. Evaluation of Ebola Virus Inhibitors for Drug Repurposing. *ACS Infect Dis*. 2015;1(7):317-26. <https://doi.org/10.1021/acsinfectdis.5b00030>
127. Zhao Y, Ren J, Harlos K, Jones DM, Zeltina A, Bowden TA, et al. Tor-emifene interacts with and destabilizes the Ebola virus glycoprotein. *Nature*. 2016;535(7610):169-72. <https://doi.org/10.1038/nature18615>