

Use of hydrophilic and hydrophobic polymers for the development of controlled release tizanidine matrix tablets

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The aim of the present study was to develop tizanidine controlled release matrix. Formulations were designed using central composite method with the help of design expert version 7.0 software. Avicel pH 101 in the range of 14-50% was used as a filler, while HPMC K4M and K100M in the range of 25-55%, Ethylcellulose 10 ST and 10FP in the range of 15 - 45% and Kollidon SR in the range of 25-60% were used as controlled release agents in designing different formulations. Various physical parameters including powder flow for blends and weight variation, thickness, hardness, friability, disintegration time and *in-vitro* release were tested for tablets. Assay of tablets were also performed as specified in USP 35 NF 32. Physical parameters of both powder blend and compressed tablets such as compressibility index, angle of repose, weight variation, thickness, hardness, friability, disintegration time and assay were evaluated and found to be satisfactory for formulations K4M2, K4M3, K4M9, K100M2, K100M3, K100M9, E10FP2, E10FP9, KSR2, KSR3 & KSR9. *In vitro* dissolution study was conducted in 900 ml of 0.1N HCl, phosphate buffer pH 4.5 and 6.8 medium using USP Apparatus II. *In vitro* release profiles indicated that formulations prepared with Ethocel 10 standard were unable to control the release of drug while formulations K4M2, K100M9, E10FP2 & KSR2 having polymer content ranging from 40-55% showed a controlled drug release pattern in the above mentioned medium. Zero-order drug release kinetics was observed for formulations K4M2, K100M9, E10FP2 & KSR2. Similarity test (f_2) results for K4M2, E10FP2 & KSR2 were found to be comparable with reference formulation K100M9. Response Surface plots were also prepared for evaluating the effect of independent variable on the responses. Stability study was performed as per ICH guidelines and the calculated shelf life was 24-30 months for formulation K4M2, K100M9 and E10FP2.

Uniterms: Tizanidine/controlled release. Hydroxypropyl methylcellulose/controlled release agent. Ethylcellulose/controlled release agent. Kollidon SR/controlled release agent. Tablets/controlled release.

O objetivo do presente estudo foi desenvolver matriz de de tizanidina de liberação controlada. As formulações foram projetadas usando o método do componente, central com a ajuda de *software* Design expert®, versão 7.0. Utilizou-se Avicel pH 101, no intervalo de 14-50%, como material de preenchimento, enquanto HPMC K4M e K100M, no intervalo de 25-55%, Etilcelulose 10 ST e 10FP, no intervalo de 15-45% e Kollidon SR, na faixa de 25-60% foram utilizados como agentes de liberação controlada, no planejamento de formulações diferentes. Vários parâmetros físicos, incluindo o fluxo de pó para as misturas e variação de peso, espessura, dureza, friabilidade, tempo de desintegração e liberação *in vitro*, foram testados para comprimidos. Ensaio dos comprimidos foram, também, realizados, tal como especificado em USP 35 NF 32. Avaliaram-se os parâmetros físicos de ambos, mistura em pó e comprimidos, como índice de compressibilidade, ângulo de repouso, variação de peso, espessura, dureza, friabilidade, tempo de desintegração e de ensaio, considerando-os satisfatórios para as formulações K4M2, K4M3, K4M9, K100M2, K100M3, K100M9, E10FP2, E10FP9, KSR2, KSR3 e KSR9. O estudo de dissolução *in vitro* foi realizado em 900 mL de HCl 0,1 N, tampão de fosfato pH 4,5 e meio 6,8, usando

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aparelho USP II. Os perfis de liberação *in vitro* indicaram que as formulações preparadas com Ethocel 10 padrão não foram capazes de controlar a liberação do fármaco, enquanto as formulações K4M2, K100M9, E10FP2 e KSR2, com teor de polímero variando entre 40 e 55% apresentaram padrão de liberação controlada de fármaco no meio anteriormente mencionado. Observou-se cinética de liberação de fármaco de ordem zero para as formulações K4M2, K100M9, E10FP2 e KSR2. Resultados do teste de similaridade (f_2) para K4M2, E10FP2 e KSR2 foram comparáveis com a formulação de referência K100M9. Gráficos de superfície de resposta também avaliaram o efeito da variável independente sobre as respostas. Estudo de estabilidade foi realizado conforme as diretrizes do ICH e a vida de prateleira calculada foi de 24-30 meses para as formulações K4M2, K100M9 e E10FP2.

Unitermos: Tizanidina/liberação controlada. Hidroxipropilmetilcelulose/agente de liberação controlada. Etilcelulose/agente de liberação controlada. Kollidon SR./agente de liberação controlada. Comprimidos/liberação controlada.

INTRODUCTION

Controlled release formulations of drugs may increase their therapeutic benefits, minimize side effects and enhance the patient compliance, therefore the overall disease management will be improved. Matrix based drug release systems offer many advantages such as the combination of matrix former and other materials can help in correlating the release characteristics to the requirements of drug and disease condition (Wilson, Crowley, 2011).

Tizanidine hydrochloride is an imidazoline derivative. It is a white to off white fine crystalline and odorless powder which is slightly soluble in water and methanol. Its solubility in water decreases with the increase in pH (Moffat, Osselton, Widdop, 2011). Acts on α_2 receptors that are centrally located and produces a myotonolytic response on skeletal muscle (Wagstaff, Bryson, 1997) It acts mainly at spinal and supraspinal levels to inhibit excitatory inter neuron and is used for the symptomatic relief of spasticity associated with multiple sclerosis or with spinal cord injury or disease (Sweetman, 2009). It is also used in relieving pain with many disorders like myofascial (Meythaler *et al.*, 2001), refractory and neuropathic pain, chronic tension type headache and chronic daily headache (Saper *et al.*, 2002). After oral administration, tizanidine is widely absorbed from the gastrointestinal tract (53 to 66%). Peak plasma concentration is achieved within 1 to 2 h. It has a bioavailability of about 34% to 40% and has a half-life of 2.5 h. Protein binding of tizanidine is 30% and it undergoes rapid and extensive first-pass metabolism in the liver (approximately 95% of a dose) (Shanker *et al.*, 2009). The therapeutic dose of tizanidine is 2 mg and 4 mg twice a day. The maximum recommended dose is 36 mg/day (Kulkarni, Babu, 2012). The short half-life, low bioavailability and extensive first-pass metabolism make tizanidine a good candidate for the development of controlled release formulations.

In the present study controlled release formulations of tizanidine hydrochloride were designed using different viscosity grades of hydroxypropyl methylcellulose, ethyl cellulose and Kollidon® SR. HPMC is an important hydrophilic polymer extensively used for controlled release dosage forms development (Siepmann, Streubel, Peppas, 2002). Its fast gel forming characteristics not only control the initial release but also exert sustained release effect through strong viscous gel formation (Reza, Quadir, Haider, 2003). Moreover, its non-toxic nature, easy compressibility made it ideal for controlled release formulations of many drugs (Shoaib *et al.*, 2010). Ethyl cellulose is also a nontoxic, inert hydrophobic polymer widely used in sustained release formulations and to control the dissolution rate of drugs (Quadir *et al.*, 2005). As a matrix forming excipient for controlled release formulations Kollidon® SR is also an extensively used polymer consisting of 80% water insoluble poly(vinyl acetate) with a molecular weight of about 450.000 and about 20% water soluble poly(vinyl pyrrolidone), named as Kollidon® 30 (Strübing, Metz, Mäder, 2008). Many researchers (Draganoiu, Andheria, Sakr, 2001; Shao *et al.*, 2001) used Kollidon® SR to delay the release of highly water soluble drugs such as propranolol HCl, diphenhydramine HCl etc., with low friability and high crushing strengths at low compression pressure (Kolter, 2002).

Response surface methodology applied for illustration of effect of independent variables (such as polymers and fillers) on the dependent variables or responses (such as disintegration time and drug release). Response surface methodology (RSM) is a statistical tool for optimization of multifactor experiments (Hamsaveni, Prapulla, Divakar, 2001; Chiang, Chang, Shieh, 2003; Zhang *et al.*, 2007). It is used to illustrate the correlation between independent variables and responses (Vicente *et al.*, 1998). RSM require less effort with minimum number of trials as compare to other approaches (Liyana-

Pathirana, Shahidi, 2005; Xiong *et al.*, 2009). Central composite rotatable design (CCRD), is one of the design approaches in RSM, was developed by Box and Wilson (Box, Wilson, 1951) and later improvement has been made in this design by Box and Hunter (Box, Hunter, 1957). CCRD is an effective optimization technique which enables to recognize optimum responses around center points through its rotatable characteristics (Zhang *et al.*, 2010 a).

The aim of present work is to develop controlled release tizanidine matrix tablets using different polymers i.e., HPMC, Ethocel and Kollidon® SR, by direct compression method. Formulation of controlled release tizanidine tablets with these polymers using software Design expert® and application of response surface models, has not been reported earlier. Study showed importance and give a detailed analysis of work which proved formulation of tizanidine with hydrophilic and hydrophobic polymers.

MATERIAL AND METHODS

Material

The following materials were used: tizanidine hydrochloride (Novartis Pharma, Karachi), HPMC K4M & K100M, Ethocel 10 Standard Premium and Ethocel 10 FP (Colorcon, England), Kollidon SR (BASF, Germany) Avicel PH 101 (FMC Corporation, USA) and magnesium stearate (Dow Chemical, USA).

Methods

Calculation of dose for controlled release tizanidine tablets

The total dose for tizanidine controlled release tablet can be calculated by using the following equation (Shoab *et al.*, 2010).

$$D_t = D_n \left(1 + 0.693 \times T_d / t_{1/2} \right) \quad \text{Eq (1)}$$

where D_t is the prolonged action dose, D_n is the normal dose and T_d is a required maintenance time.

For tizanidine hydrochloride, immediate release dose (D_n) is 4mg and half-life is 2.5 hours. Therefore, for a controlled release during 24h (T_d), 30 mg of tizanidine (equivalent to 34 mg of tizanidine hydrochloride) per tablet was taken.

Preparation of tablets

Formulations were designed by using a software

Design expert®. Different formulations of HPMC viscosity grades of K4M & K100M, Ethocel 10 ST (standard grade) & 10 FP (fine particle grade) and Kollidon SR were prepared by direct compression method. Each tablet containing 34 mg of tizanidine hydrochloride (equivalent to 30 mg of tizanidine) and 2mg of magnesium stearate, while polymers HPMC K4M & K100M ranging from 25 to 55%, Ethocel 10ST & 10 FP (15% to 45%) and Kollidon SR (25 to 60%) were used as independent variable (X1) and Avicel PH 101 (14 to 50% in HPMC formulations, 26 to 48% in Ethocel formulations and 8-51% in Kollidon SR formulations) was used as second independent variable (X2). Formulation excipients and active ingredient were passed through 40-mesh size sieve and accurately weighed. Excipients and active ingredient of each formulation were blended by tumbling method in a polybag for about 8-10 min and then compressed on manually operated single punch tablet press (Korsch Erweka, Frankfurt, Germany). Convex round shaped punch was used and tablets were pressed in a range of 96 to 126 mg.

Experimental design

To find out the optimum level of variable, a two factor with five levels CCD was developed. It consists of factorial points at two levels, axial points at two levels and center point. Hence, independent variables (X1: Polymer and X2: Avicel PH 101) at five levels were considered. The levels were $-\alpha$, -1 , 0 , 1 and $+\alpha$. The alpha value (1.414) was taken to execute design rotatability. Disintegration time and drug release both at 2 and 8 h were selected as responses. The coded and actual values of variables are given in Table I. By using Design Expert software (Version 7, Stat-Ease Inc., Minneapolis, MN), CCD generated a total of nine (9) experiments with four factorial, four axial and one center point. Hao *et al* also used central composite response surface design with five replicated center points (Hao *et al.*, 2012).

Micromeritic Study

Micromeritic properties of powder blends such as tapped density, bulk density, Hausner's ratio, compressibility index and angle of repose were evaluated using the procedure specified in US pharmacopeia (USP35-NF30, 2012).

Hausner's ratio, compressibility index and angle of repose were calculated by the following equations.

$$\text{Hausner's ratio} = (\rho_{\text{tapped}} / \rho_{\text{bulk}}) \quad \text{Eq (2)}$$

$$\text{Compressibility Index} = [100 \times (\rho_{\text{tapped}} - \rho_{\text{bulk}}) / \rho_{\text{tapped}}] \quad \text{Eq (3)}$$

TABLE I - A two factor central composite rotatable design of experiments for tizanidine formulations

Formulation Code	Coded factor level		Factors Amount (%)		Factors Amount (mg)		Total Wt. of Tablet (mg)	Total wt. adjusted (mg)
	X 1	X 2						
<i>HPMC K4M</i>	<i>HPMC K4M</i>	<i>Avicel PH 101</i>	<i>HPMC K4M</i>	<i>Avicel PH 101</i>	<i>HPMC K4M</i>	<i>Avicel PH 101</i>		
K4M 1	-1	-1	30.00	20.00	30.84	20.56	87.40	88
K4M 2	1	-1	50.00	20.00	51.40	20.56	107.96	108
K4M 3	-1	1	30.00	45.00	30.84	46.26	113.10	113
K4M 4	1	1	50.00	45.00	51.40	46.26	133.66	134
K4M 5	-1.414	0	25.86	32.50	26.58	33.41	95.99	96
K4M 6	1.414	0	54.14	32.50	55.66	33.41	125.07	125
K4M 7	0	-1.414	40.00	14.82	41.12	15.23	92.35	93
K4M 8	0	1.414	40.00	50.18	41.12	51.58	128.70	128
K4M 9	0	0	40.00	32.50	41.12	33.41	110.53	110
<i>HPMC K100M</i>	<i>HPMC K100M</i>	<i>Avicel PH 101</i>	<i>HPMC K100M</i>	<i>Avicel PH 101</i>	<i>HPMC K100M</i>	<i>Avicel PH 101</i>		
K100M 1	-1	-1	30.00	20.00	30.84	20.56	87.40	88
K100M 2	1	-1	50.00	20.00	51.40	20.56	107.96	108
K100M 3	-1	1	30.00	45.00	30.84	46.26	113.10	113
K100M 4	1	1	50.00	45.00	51.40	46.26	133.66	134
K100M 5	-1.414	0	25.86	32.50	26.58	33.41	95.99	96
K100M 6	1.414	0	54.14	32.50	55.66	33.41	125.07	125
K100M 7	0	-1.414	40.00	14.82	41.12	15.23	92.35	93
K100M 8	0	1.414	40.00	50.18	41.12	51.58	128.70	128
K100M 9	0	0	40.00	32.50	41.12	33.41	110.53	110
<i>Ethocel 10 Standard</i>	<i>Ethocel 10 ST</i>	<i>Avicel PH 101</i>	<i>Ethocel 10 ST</i>	<i>Avicel PH 101</i>	<i>Ethocel 10 ST</i>	<i>Avicel PH 101</i>		
E10ST 1	-1	-1	20.00	30.00	22.00	33.00	91.00	91
E10ST 2	1	-1	40.00	30.00	44.00	33.00	113.00	113
E10ST 3	-1	1	20.00	45.00	22.00	49.50	107.50	108
E10ST 4	1	1	40.00	45.00	44.00	49.50	129.50	130
E10ST 5	-1.414	0	15.86	37.50	17.45	41.25	94.70	95
E10ST 6	1.414	0	44.14	37.50	48.56	41.25	125.81	126
E10ST 7	0	-1.414	30.00	26.89	33.00	29.58	98.58	99
E10ST 8	0	1.414	30.00	48.11	33.00	52.92	121.92	122
E10ST 9	0	0	30.00	37.50	33.00	41.25	110.25	110
<i>Ethocel 10 FP</i>	<i>Ethocel 10 FP</i>	<i>Avicel PH 101</i>	<i>Ethocel 10 FP</i>	<i>Avicel PH 101</i>	<i>Ethocel 10 FP</i>	<i>Avicel PH 101</i>		
E10FP 1	-1	-1	20.00	30.00	22.00	33.00	91.00	91
E10FP 2	1	-1	40.00	30.00	44.00	33.00	113.00	113
E10FP 3	-1	1	20.00	45.00	22.00	49.50	107.50	108
E10FP 4	1	1	40.00	45.00	44.00	49.50	129.50	130
E10FP 5	-1.414	0	15.86	37.50	17.45	41.25	94.70	95
E10FP 6	1.414	0	44.14	37.50	48.56	41.25	125.81	126
E10FP 7	0	-1.414	30.00	26.89	33.00	29.58	98.58	99
E10FP 8	0	1.414	30.00	48.11	33.00	52.92	121.92	122
E10FP 9	0	0	30.00	37.50	33.00	41.25	110.25	110
<i>Kollidon SR</i>	<i>KSR</i>	<i>Avicel PH 101</i>	<i>KSR</i>	<i>Avicel PH 101</i>	<i>KSR</i>	<i>Avicel PH 101</i>		
KSR 1	-1	-1	30.00	15.00	33.00	16.50	85.50	86
KSR 2	1	-1	55.00	15.00	60.50	16.50	113.00	113
KSR 3	-1	1	30.00	45.00	33.00	49.50	118.50	118
KSR 4	1	1	55.00	45.00	60.50	49.50	146.00	146
KSR 5	-1.414	0	24.82	30.00	27.30	33.00	96.30	96
KSR 6	1.414	0	60.18	30.00	66.20	33.00	135.20	135
KSR 7	0	-1.414	42.50	8.79	46.75	9.67	92.42	92
KSR 8	0	1.414	42.50	51.21	46.75	56.33	139.08	139
KSR 9	0	0	42.50	30.00	46.75	33.00	115.75	116

$$\tan(\alpha) = \text{height} / 0.5 \text{ base} \quad \text{Eq (4)}$$

where ρ_{tapped} and ρ_{bulk} were the tapped and bulk densities of blends, respectively and α was the angle of repose.

Evaluation of tizanidine tablets

Tizanidine tablets were evaluated using several physical parameters including weight variation, hardness, thickness, friability and disintegration. Weight variation was performed by using digital balance (Sartorius CP 224S, Germany) for all compressed formulations. Tablet hardness was determined using Fujiwara Seisakusho tablet hardness tester, Ogawa Seiko Co Ltd, Tokyo, Japan. Tablet disintegration test were performed using USP basket rack assembly (Erweka disintegration tester, ZT2, Heusenstamm, Germany). Tablet friability was carried out using Roche type friabilator (H Jurgens friabilator GmbH & Co. D-2800 Bremen, Germany) (USP 35-NF 30, 2012).

Swelling studies

Swelling studies of tablets were performed by gently placing a tablet (using a wire) in a beaker containing about 250 mL of distilled water at room temperature. The weights of swollen tablets (after absorbing excess water through a filter paper) were recorded at 1, 2, 3, 4, 6 and 8 h. Water uptake (%) was expressed as a percentage of initial tablet weight (Cao *et al.*, 2005).

$$\% S = \frac{W_t - W_0}{W_0} \times 100 \quad \text{Eq (5)}$$

where S is the swelling of tablet, W_t is the weight of swollen tablet and W_0 is the initial weight of tablet.

Dissolution studies

Dissolution studies were performed by placing six tablets of each formulation in 900 mL of dissolution medium at 37 ± 0.5 °C using a USP apparatus II dissolution tester (DT 600, Erweka, Germany). Paddles were rotated at 100rpm and the medium used was 0.1 N HCl, Phosphate buffer at pH 4.5 & 6.8. Samples were analyzed using UV-VIS spectrophotometer (UV-1800, Shimadzu, Kyoto, Japan) at 320 nm by withdrawing 5 mL of aliquots at regular time interval of 30min, 1, 2, 3, 4, 5, 6, 8, 10, 12, 16, 20 and 24h and filtered using a 0.45 μm filter. These aliquots were replaced by the same medium previously maintained at 37 ± 0.5 °C.

Quantification of tizanidine in the formulation

Drug quantification was performed using high performance liquid chromatographic method. The mobile

phase was composed of acetonitrile and buffer solution of sodium 1-pentanesulfonate in a ratio of 4:1 (buffer solution was prepared by adding 3.5g of sodium 1-pentane sulfonate in 1 liter of water and pH was adjusted with phosphoric acid solution or 1 N sodium hydroxide at pH 3.0 ± 0.05). The HPLC system consists of a pump LC-10 AT VP, Communication Bus Module CBM 102, and a UV detector SPD 10-A VP (Shimadzu Corp, Tokyo, Japan). The column used was Intersil ODS-3, 4.6 x 250mm which was maintained at 50 °C in a column oven (CTO-10A, Shimadzu Corp., Kyoto, Japan) with a flow rate of 1 mL/min. Injection volume was 10 μL and detection wavelength was 230 nm. Software used was class GC 10 ver. 2.0 (1993-2000), (Shimadzu Corp., Kyoto, Japan) was used for data processing (USP 35-NF 30, 2012).

Stability studies

The optimized formulations were then subjected to accelerated stability study as per ICH guidelines i.e. by placing tablets in a Stability chamber ((Binder GMBH Bergster, Tullingen, Germany) at 40 ± 2 °C and $75 \pm 5\%$ RH (ICH, 2003). The samples were removed and tested at 0, 1, 3 and 6 months and different physico-chemical parameters like hardness, friability, disintegration, dissolution and quantification were assessed. The shelf life was calculated using R-Gui version 2.15.2 (stab) package (The R Foundation for Statistical Computing).

Data Analysis

- Model-dependent methods

In order to propose mechanism of drug release from these formulations, *in vitro* dissolution profiles data were fitted into different kinetic models. These models were zero order (cumulative amount of drug released vs time), first-order (log cumulative percentage of drug remaining vs time), Higuchi's (cumulative percentage of drug released vs square root of time), Hixson-Crowell (cube root percent drug remaining vs time) and Korsmeyer's (log cumulative percentage of drug released vs log time).

- Zero order equation:

$$Q_t = K_0 t \quad \text{Eq (6)}$$

where K_0 is the zero-order rate constant expressed in units of concentration/time, t is the time in hours, and Q_t is the amount of drug release in time t.

- First-order equation:

$$\text{Log} Q_t = \text{log} Q_0 - k_t / 2.303 \quad \text{Eq (7)}$$

where Q_t is the amount released at time t , Q_0 is the initial amount of drug in solution and k is the first order rate constant and t is the time.

- Higuchi's equation:

$$Q = kt^{1/2} \quad \text{Eq (8)}$$

where k is the release rate constant and t is the time in hours. Hence, the drug release rate is proportional to the reciprocal of the square root of time (David, 2002).

- Hixson–Crowell Cube Root equation

$$Q_0^{1/3} - Q_t^{1/3} = K_{HC} \times t \quad \text{Eq (9)}$$

where K_{HC} is the Hixson–Crowell rate constant, Q_0 is the initial amount of drug and Q_t is the amount of drug release at time t (Higuchi, 1963).

For zero order, Higuchi, and Hixson–Crowell model, the rate constant were also calculated that is simply equal to the slope of the straight line:

$$K_0 = \text{Slope} \quad \text{Eq (10)}$$

Below is showed the equation used for first-order rate constant

$$K = -\text{Slope} \times 2.303 \quad \text{Eq (11)}$$

Korsmeyer's equation (Power law),

$$M_t / M_\infty = Kt^n \quad \text{Eq (12)}$$

where M_t is the amount of drug released at time t , M_∞ is the amount of drug released after infinite time (total drug in a dosage form), K is the Korsmeyer's dissolution rate constant and n is the release exponent which was calculated through the slope of the straight line which characterizes the mechanism of release (Hixson, Crowell, 1931).

For matrix devices that are cylindrical-shaped, if the exponent n value is 0.45 it is indicative of Fickian release (case I), for non-Fickian release (anomalous) it should be >0.45 but <0.89 , value of 0.89 is indicative of case II (zero order) release, and >0.89 is super case II type of release (Korsmeyer *et al.*, 1983).

Model Independent Method

- Similarity Factor (f_2)

The similarity factor (f_2) is a logarithmic reciprocal

square root transformation of the sum of squared error and is a measurement of the similarity in the percent dissolution between the two curves:

$$f_2 = 50 \times \log \left\{ \left[1 + \left(\frac{1}{N} \right) \sum (R_i - T_i)^2 \right]^{-0.5} \right\} \times 100 \quad \text{Eq (13)}$$

where R_i is the percent dissolved of referenced drug, T_i is the percent dissolved of test drug at each time point and N is the number of samples. Its value will be 100 where test and reference drug profiles are identical. Dissimilarity in profiles increases with the decrease in f_2 (Costa, Sousa Lobo, 2001).

RESULTS AND DISCUSSION

Formulation of tizanidine matrix tablets

Tizanidine controlled release formulations were designed using software Design Expert version 7.0. The coded and actual quantities of individual content of tablet for different polymers i.e HPMC (K4M & K100M) Ethocel (10 ST & 10 FP) and Kollidon SR are shown in Table I. HPMC K4M and K100M formulations were assigned formulation code K4M and K100M as prefix followed by numeric number as suffix. Similarly, formulations containing Ethocel 10 Standard grade, Ethocel 10 fine particle grade and Kollidon SR were assigned formulation codes E10ST, E10FP and KSR, respectively and followed by a numeric number as suffix.

Evaluation of powder blends and tablets

Micromeritic evaluation of powder blends, Hausner's ratio, compressibility index and angle of repose were calculated for the formulations and showed in Table II. The powder blends which comply with USP standard and categorized as Fair to Excellent, were chosen for compression and further studies. The formulations which complied with USP standards in terms of flow properties were K4M2, K4M3, K4M5, K4M9, K100M2, K100M3, K100M5, K100M9, E10ST2, E10ST6, E10ST9, E10FP2, E10FP3, E10FP9, KSR2, KSR3, KSR5 and KSR9. These formulations were evaluated for physical parameters such as thickness, hardness, weight variation, friability, disintegration time and assay (Table III). The mean thickness and hardness of tablets were in the range of 3.67 ± 0.14 to 4.39 ± 0.18 mm and 3.40 ± 0.78 to 12.92 ± 1.80 kg, respectively. Powder flowability and level of powder in dye often regulate variation in tablet thickness (Davies, 1995). The hardness is an attribute

due to binding characteristic of the filler and the force of compression with which the ingredients have been compressed (Siddiqui, Nazzal, 2007). Weight variation was in the range of 95.74 ± 6.32 to 125.35 ± 4.53 mg. Friability of compressed formulations was $< 1\%$ except

for formulations K4M5 and KSR5. Disintegration time of less than 4 hours was observed for K4M5, K100M5, E10ST 2, 6 & 9, E10FP3 and KSR 5 formulations. Early disintegration observed in formulation where the concentration of polymers was low, it seemed that low

TABLE II - Micromeritic Properties of different formulation blends

Formulation code	Mass	Bulk Volume	Tapped volume	Bulk density	Tapped density	Haunser Ratio	Compressibility Index	Angle of repose	Flow properties according to
	(g)	(mL)	(mL)	(g/mL)	(g/mL)		%	θ	USP 35
K4M 1	10.00	17.00	12.00	0.59	0.83	1.42	29.41	53.28	Poor
K4M 2	10.00	17.00	16.00	0.59	0.63	1.06	5.88	29.56	Excellent
K4M 3	10.00	19.00	16.00	0.53	0.63	1.19	15.79	36.32	Fair
K4M 4	10.00	20.00	14.00	0.50	0.71	1.43	30.00	49.83	Poor
K4M 5	10.00	20.00	16.00	0.50	0.63	1.25	20.00	38.39	Fair
K4M 6	10.00	20.00	14.00	0.50	0.71	1.43	30.00	53.99	Poor
K4M 7	10.00	17.00	12.00	0.59	0.83	1.42	29.41	54.21	Poor
K4M 8	10.00	18.00	12.00	0.56	0.83	1.50	33.33	58.18	V Poor
K4M 9	10.00	17.00	15.00	0.59	0.67	1.13	11.76	33.20	Good
K100M 1	10.00	17.00	11.00	0.59	0.91	1.55	35.29	60.79	V Poor
K100M 2	10.00	18.00	16.00	0.56	0.63	1.13	11.11	32.11	Good
K100M 3	10.00	20.00	17.00	0.50	0.59	1.18	15.00	33.28	Good
K100M 4	10.00	19.00	11.00	0.53	0.91	1.73	42.11	70.45	V V Poor
K100M 5	10.00	17.00	13.00	0.59	0.77	1.31	23.53	38.12	Fair
K100M 6	10.00	20.00	15.00	0.50	0.67	1.33	25.00	42.66	Passable
K100M 7	10.00	17.00	12.00	0.59	0.83	1.42	29.41	46.32	Poor
K100M 8	10.00	17.00	11.00	0.59	0.91	1.55	35.29	60.28	V Poor
K100M 9	10.00	18.00	17.00	0.56	0.59	1.06	5.56	27.19	Excellent
E10ST 1	10.00	24.00	16.00	0.42	0.63	1.50	33.33	61.24	V Poor
E10ST 2	10.00	23.00	19.00	0.43	0.53	1.21	17.39	38.13	Fair
E10ST 3	10.00	23.00	17.00	0.43	0.59	1.35	26.09	51.22	Poor
E10ST 4	10.00	22.00	15.00	0.45	0.67	1.47	31.82	58.54	V Poor
E10ST 5	10.00	23.00	15.00	0.43	0.67	1.53	34.78	64.87	V Poor
E10ST 6	10.00	24.00	21.00	0.42	0.48	1.14	12.50	34.25	Good
E10ST 7	10.00	24.00	18.00	0.42	0.56	1.33	25.00	43.30	Passable
E10ST 8	10.00	21.00	15.00	0.48	0.67	1.40	28.57	48.74	Poor
E10ST 9	10.00	22.00	18.00	0.45	0.56	1.22	18.18	37.34	Fair
E10FP 1	10.00	27.00	21.00	0.37	0.48	1.29	22.22	43.23	Passable
E10FP 2	10.00	28.00	24.00	0.36	0.42	1.17	14.29	32.12	Good
E10FP 3	10.00	29.00	24.00	0.34	0.42	1.21	17.24	37.75	Fair
E10FP 4	10.00	31.00	24.00	0.32	0.42	1.29	22.58	44.09	Passable
E10FP 5	10.00	31.00	22.00	0.32	0.45	1.41	29.03	53.37	Poor
E10FP 6	10.00	30.00	20.00	0.33	0.50	1.50	33.33	58.16	V Poor
E10FP 7	10.00	28.00	18.00	0.36	0.56	1.56	35.71	62.97	V Poor
E10FP 8	10.00	29.00	22.00	0.34	0.45	1.32	24.14	44.23	Passable
E10FP 9	10.00	31.00	26.00	0.32	0.38	1.19	16.13	37.32	Fair
KSR 1	10.00	19.00	14.00	0.53	0.71	1.36	26.32	51.55	Poor
KSR 2	10.00	19.00	18.00	0.53	0.56	1.06	5.26	27.19	Excellent
KSR 3	10.00	21.00	18.00	0.48	0.56	1.17	14.29	34.31	Good
KSR 4	10.00	22.00	16.00	0.45	0.63	1.38	27.27	47.5	Poor
KSR 5	10.00	18.00	15.00	0.56	0.67	1.20	16.67	39.25	Fair
KSR 6	10.00	22.00	15.00	0.45	0.67	1.47	31.82	57.86	V Poor
KSR 7	10.00	20.00	14.00	0.50	0.71	1.43	30.00	46.82	Poor
KSR 8	10.00	22.00	17.00	0.45	0.59	1.29	22.73	42.51	Passable
KSR 9	10.00	20.00	17.00	0.50	0.59	1.18	15.00	33.62	Good

TABLE III - Physical Evaluation and assay of tizanidine matrix

Formulation code	Thickness (mm)	Hardness (kg)	Weight Variation (mg)	Friability (%)	Disintegration time (hrs)	Assay (%)
K4M 2	3.96 ± 0.10	4.81 ± 0.52	108.23 ± 2.80	0.76	4.82	98.76
K4M 3	4.05 ± 0.12	4.20 ± 0.38	113.21 ± 2.80	0.91	4.18	97.25
K4M 5	3.67 ± 0.14	3.74 ± 0.70	96.98 ± 5.00	1.14	3.55	96.38
K4M 9	3.99 ± 0.12	4.69 ± 1.18	110.71 ± 4.06	0.94	4.30	98.05
K100M 2	4.01 ± 0.14	5.87 ± 0.68	109.30 ± 3.66	0.57	6.87	98.37
K100M 3	4.03 ± 0.16	4.30 ± 0.70	113.37 ± 4.78	0.85	4.95	97.82
K100M 5	3.97 ± 0.16	4.04 ± 1.12	95.74 ± 6.32	0.99	3.73	98.61
K100M 9	3.98 ± 0.08	5.57 ± 0.76	110.27 ± 4.00	0.63	6.58	98.87
E10ST 2	4.01 ± 0.15	8.04 ± 1.86	113.54 ± 3.71	0.76	1.42	96.81
E10ST 6	4.39 ± 0.18	8.53 ± 2.26	125.35 ± 4.53	0.69	2.10	97.32
E19ST 9	4.05 ± 0.13	7.96 ± 1.57	109.85 ± 3.82	0.97	1.38	96.18
E10FP 2	3.99 ± 0.08	12.92 ± 1.80	113.77 ± 3.74	0.30	5.72	98.79
E10FP 3	4.00 ± 0.10	11.75 ± 2.14	109.46 ± 4.5	0.95	3.68	96.32
E10FP 9	3.98 ± 0.14	12.57 ± 2.30	110.82 ± 5.38	0.47	5.13	98.24
KSR 2	4.24 ± 0.10	5.01 ± 0.74	113.30 ± 3.04	0.29	7.52	98.39
KSR 3	4.07 ± 0.14	4.24 ± 0.88	118.97 ± 3.98	0.84	4.25	96.51
KSR 5	4.04 ± 0.12	3.40 ± 0.78	96.82 ± 5.28	1.53	3.15	95.23
KSR 9	4.20 ± 0.12	4.20 ± 0.58	116.05 ± 4.04	0.63	6.27	97.27

concentration of polymers was unable to control tablet disintegration upto 4 hours. It was also observed that tablets formulated with Ethocel 10 Standard grade were unable to control the disintegration that may be due to their larger particle size which allows water to penetrate inside the void spaces as reported by Crowley *et al.* (2004). Assay of all compressed formulations were within the limits as specified in USP (USP35-NF30, 2012).

Study of hydration behavior

Swelling studies revealed the swelling properties of tablets and studied as a function of polymers used in the study. The swelling behavior of a polymer enables the formation of swellable layer or the gel layer. The gel layer hindered movement of drug from inner dry layer and enable the release of drug only from the upper gel layer and causing a controlled release of drug. Formulations containing hydrophilic polymers swell well while the hydrophobic polymers showed less swelling behavior.

The results of swelling behavior with different polymers were shown in Figure 1. It is clear from the results that HPMC showed highest hydration ability as compared to Kollidon SR and Ethocel due to its hydrophilic nature. Formulation containing Ethocel

showed least swelling among three tested polymers due to its higher hydrophobic nature. The data showed that by increasing the quantity of HPMC K4M and K100M, tablet swelling has been increased. Nerurkar *et al.*, also noticed that increase in concentration of polymer in matrix will cause an increase of amount of water uptake (Nerurkar *et al.*, 2005). Further, from these results it is also clear that with change of HPMC viscosity grade to higher side, swelling properties also enhanced.

In vitro drug release profiles

Tizanidine formulations release profiles in 0.1 N HCl, phosphate buffer pH 4.5 and 6.8 were presented in Figure 2, 3 and 4, respectively. Release profiles of tizanidine tablets in different dissolution media were not found significantly different. Amir and Ahmad (2010), also found the similar type of drug release behavior of tizanidine and tramadol modified release microparticles in 0.1 N HCl and pH 6.8 phosphate buffer. It was found that formulations with Ethocel 10 standard grade were unable to control the release and nearly the entire drug has been released within 4 hours in the three dissolution media. According to Percolation theory, in a matrix formulation of hydrophilic drug and hydrophobic polymer, drug release

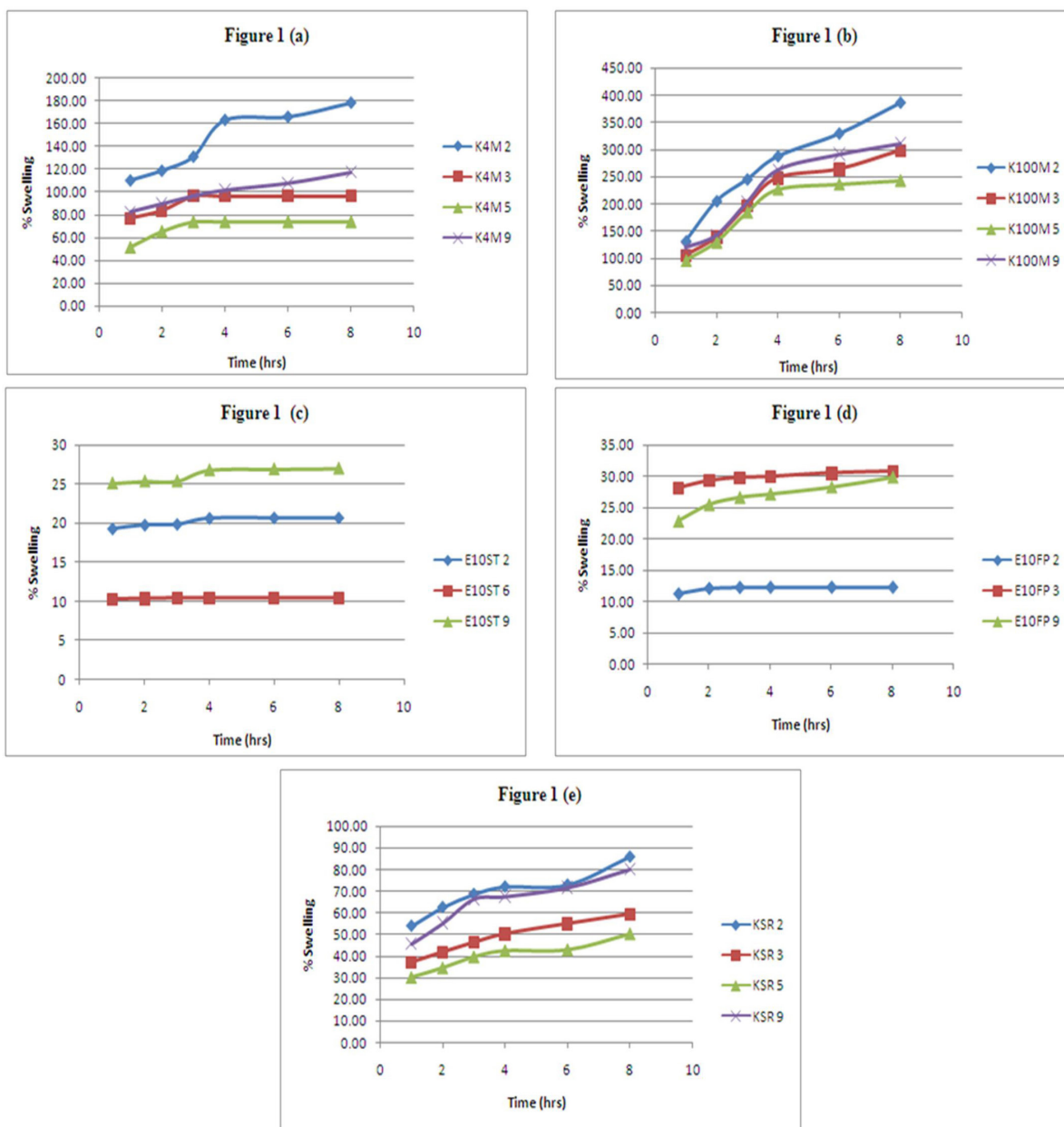


FIGURE 1 - Hydration behavior of tizanidine matrix formulations containing (a) HPMC K4M (b) HPMC K100M (c) Ethocel 10ST (d) Ethocel 10 FP (e) Kollidon SR.

was derived by dissolution of drug through capillaries composed of interconnecting drug particle cluster and pore network (Holman, Leuenberger, 1988). With more ethyl cellulose particles fewer pore networks were formed (Crowley *et al.*, 2004) as the case with fine particles and retard the release while in case of coarser particles the more pore network will be formed that was unable to control the release of drug.

Formulations containing 25% or less of polymer (K4M5, K100M5, E10FP3 and KSR5) were also unable to retard the release and more than 90% of drug was

available in dissolution media in 4 hours. Tablets with a polymer ratio of 30% (K4M3, K100M3 and KSR3) released about 80% of drug except in case of Ethocel 10 FP (E10FP9) where 76% release was observed with 30% of polymer. K100M9 (HPMC K100M: 40% polymer) given a good controlled release profile. From K100M9 formulation, around 40% drug released in 4h, 80% in 8h and 98% in 24 hours. Similar type of profiles were observed in formulations K4M2 (50% polymer), E10FP2 (40% polymer) and KSR2 (55% polymer) in which around 45 to 53% drug released in 4 hours was observed, 82 to

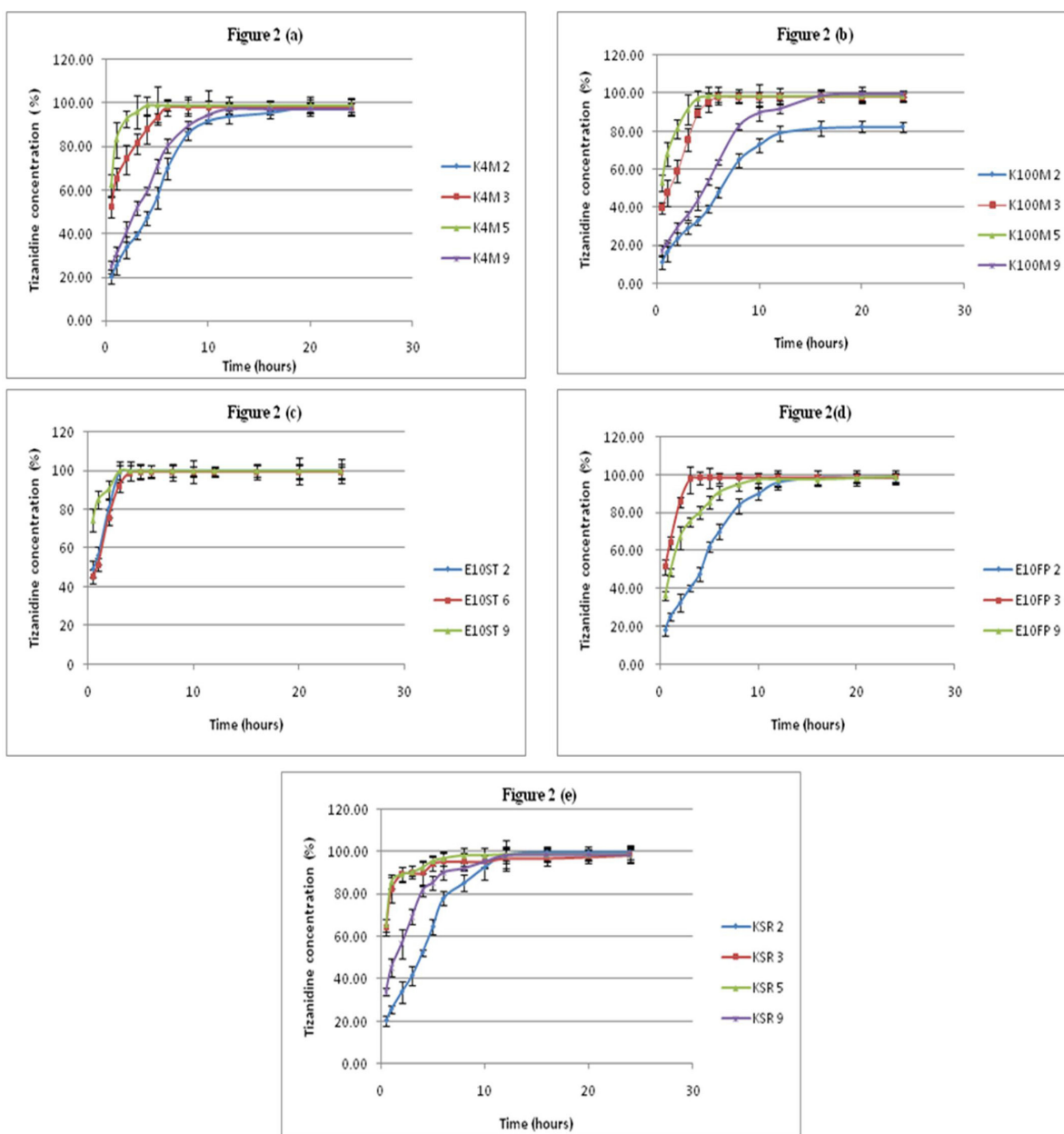


FIGURE 2 - Drug release profiles of Tizanidine formulations in 0.1 N HCl containing (a) HPMC K4M (b) HPMC K100M (c) Ethocel 10ST (d) Ethocel 10FP (e) Kollidon SR.

85% drug released in 8 hours and 94 to 99% drug released in 24 hours. It was observed in all formulations that drug release has inverse relation with polymer concentration. Barakat, Elbagory and Almurshedi (2009) found that increase in the concentration of HPMC results in reduction in the release rate from carbamazepine matrix formulation and followed non-fickian diffusion which shifted to case II with the increase in HPMC ratio in formulation, showed significant contribution through erosion. Similarly, Baviskar, Sharma and Jain (2013) observed retard release of verapamil hydrochloride matrix tablet with increase

in concentration of HPMC K15M and Eudragit RSPO. Reza, Quadir and Haider (2003) found increase in drug release rate with decreasing total polymeric content of matrix tablets of theophylline, diclofenac sodium and diltiazem hydrochloride by using plastic, hydrophobic and hydrophilic polymers. The release rate was much retarded in formulation K100M2 which contains 50% of polymer and around 80% of the drug release in 24 hours. Huang *et al.* (2004) had similar finding where release was incomplete from propranolol hydrochloride tablet containing high amount of HPMC.

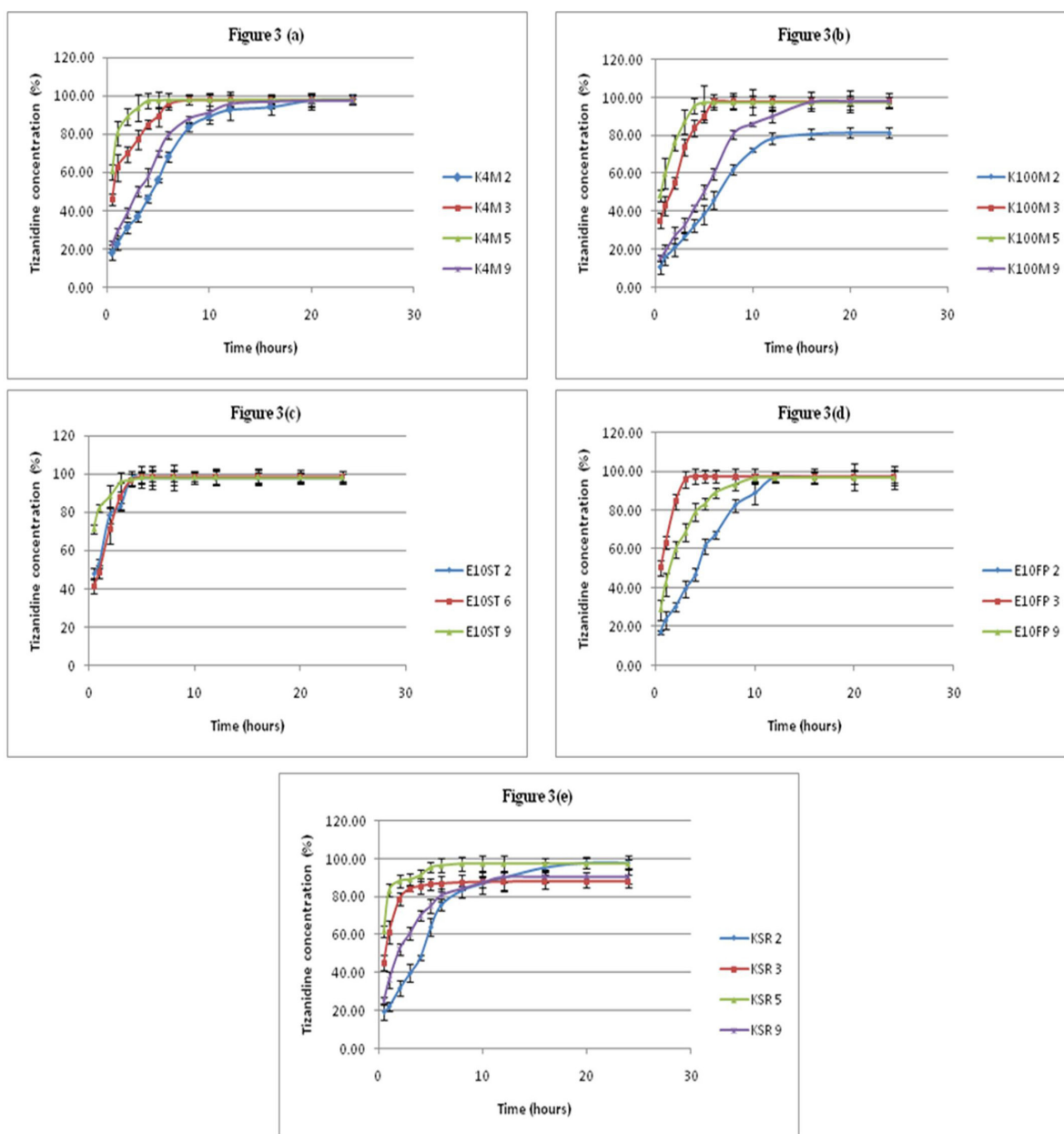


FIGURE 3 - Drug release profiles of tizanidine formulations in phosphate buffer pH 4.5 containing (a) HPMC K4M (b) HPMC K100M (c) Ethocel 10ST (d) Ethocel 10FP (e) Kollidon SR.

Drug release kinetics

Model dependent method

Drug release kinetics from tizanidine matrix tablets was described by various mathematical models and equations. The dissolution data was fitted to zero order, first order, Higuchi, Korsmeyer-Peppas and Hixon-Crowell to determine the mechanism of drug release. The Regression coefficients and release constants were calculated by DD-solver and shown in Table IV. DD Solver is an add-in program for Microsoft Excel®

for dissolution data modeling and profile comparison (Zhang *et al.*, 2010b). The dissolution data up to 8 hours were used for all formulations. The data selection was based on the data until the time which represent the dynamics of dissolution process (Polli *et al.*, 1997). The tizanidine formulations which were best support the zero order release kinetics were K4M2, K100M9, E10FP2 and KSR2 as highest linearity values were observed in 0.1 N HCl, phosphate buffer pH 4.5 and 6.8. Formulation, K100M2 although yielded zero-order but incomplete release kinetics. Bravo, Lamas and

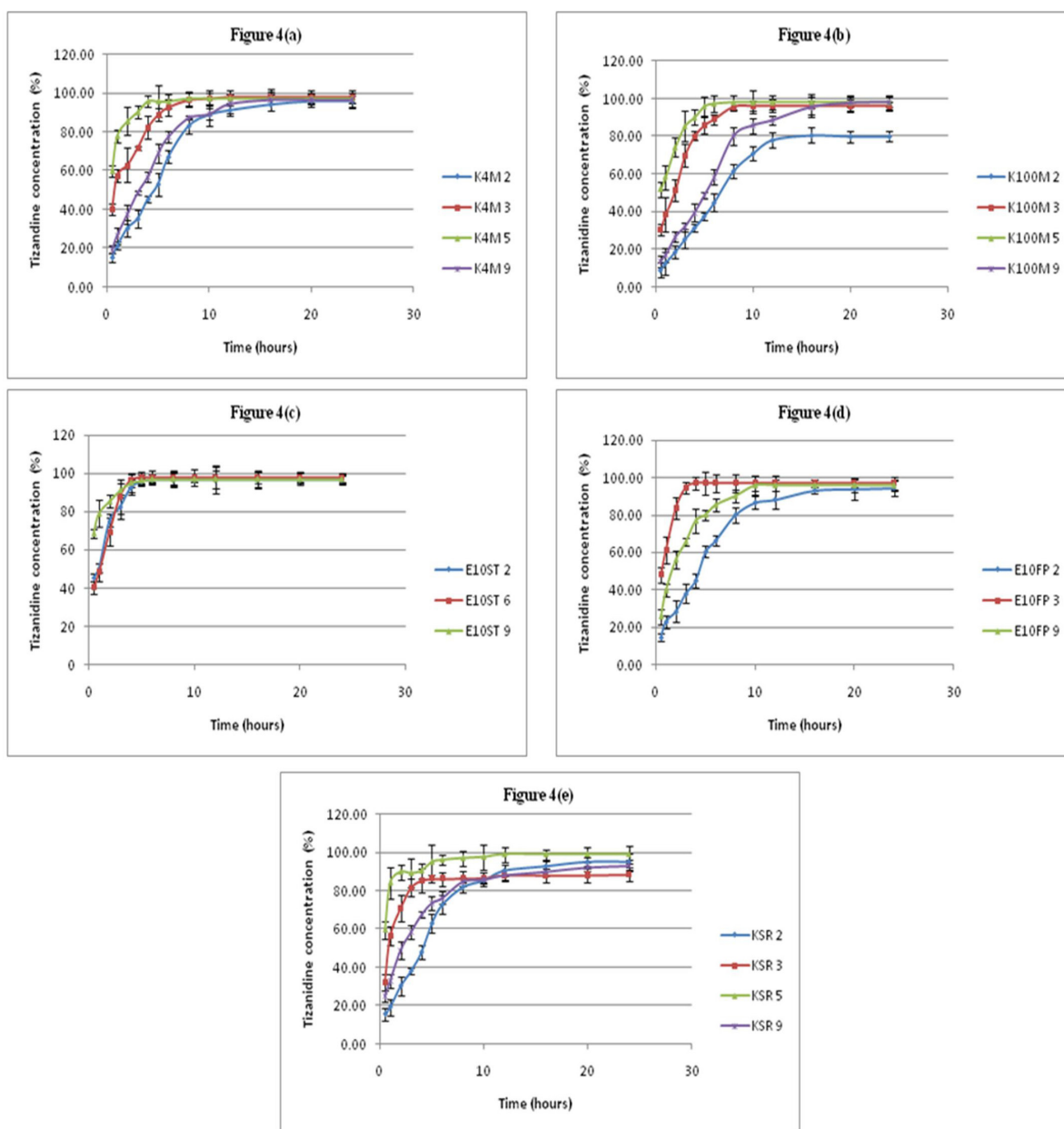


FIGURE 4 - Drug release profiles of tizanidine formulations in phosphate buffer pH 6.8 containing (a) HPMC K4M (b) HPMC K100M (c) Ethocel 10ST (d) Ethocel 10FP (e) Kollidon SR.

Salomon (2002) found zero order release kinetics for diclofenac sodium HPMC matrix tablets with highest regression coefficient values followed by Higuchi and first order. Sankalia Sankalia, Mashru (2008) have formulated glipizide matrix tablets with xanthan:MCC PH301 and xanthan:HPMC K4M:Starch 1500 with Korsmeyer–Peppas' and zero-order release mechanism, respectively. Jan *et al.* (2011) successfully developed sustained release Ketoprofen tablets with different grades of Ethocel FP and found release kinetics nearly zero-order. Sahoo *et al.* (2008) prepared controlled release

propranolol hydrochloride tablet by direct compression with 40% Kollidon SR that showed a zero-order release kinetics.

The diffusional coefficient (n) obtained from Korsmeyers equation were in the range of > 0.45 to < 0.89 for K4M2, K100M9, E10FP2 and KSR2 which showed that drug was release through anomalous transport also known as non-Fickian release that was an indication that the release of drug is controlled by both diffusion and erosion mechanism (Peppas, 1985). This equation has also been used by many researchers

TABLE IV - Model dependent assessment of tizanidine formulations

	Zero order		First order		Higuchi		Korsmeyer-Peppas			Hixon-crowell	
	r^2	$K_0(h^{-1})$	r^2	$K_1(h^{-1})$	r^2	$K_H(h^{-1/2})$	r^2	n	$K_{kp}(h^{-n})$	r^2	$K_{HC}(h^{-1/3})$
0.1 N HCl											
K4M 2	0.996	11.586	0.964	0.192	0.972	26.792	0.982	0.641	21.498	0.976	0.055
K4M 3	0.930	17.217	0.951	0.923	0.978	42.347	0.991	0.226	63.683	0.974	0.180
K4M 9	0.992	13.355	0.984	0.259	0.994	31.333	0.994	0.526	30.089	0.992	0.072
K100M 2	0.994	8.310	0.977	0.115	0.971	19.058	0.985	0.726	13.342	0.983	0.034
K100M 3	0.927	16.894	0.972	0.566	0.970	40.719	0.976	0.358	50.441	0.980	0.146
K100M 9	0.997	10.830	0.969	0.170	0.971	24.870	0.986	0.713	17.794	0.979	0.049
E10FP 2	0.996	11.653	0.978	0.195	0.983	26.959	0.990	0.642	21.589	0.987	0.056
E10FP 9	0.927	15.951	0.995	0.526	0.978	38.660	0.989	0.325	50.353	0.991	0.139
KSR 2	0.991	12.287	0.975	0.215	0.981	28.472	0.987	0.629	23.277	0.985	0.061
KSR 3	0.778	17.573	0.979	1.821	0.862	44.177	0.919	0.117	77.467	0.936	0.191
KSR 9	0.938	15.559	0.991	0.449	0.981	37.471	0.987	0.363	46.104	0.996	0.119
pH 4.5											
K4M 2	0.997	11.257	0.971	0.182	0.975	25.919	0.988	0.686	19.373	0.982	0.052
K4M 3	0.941	16.764	0.959	0.732	0.982	40.997	0.991	0.257	58.960	0.965	0.171
K4M 9	0.990	13.075	0.986	0.250	0.993	30.579	0.994	0.554	28.138	0.993	0.069
K100M 2	0.997	7.971	0.982	0.108	0.973	18.242	0.988	0.749	12.322	0.988	0.033
K100M 3	0.944	16.422	0.981	0.487	0.981	39.311	0.984	0.401	45.674	0.987	0.127
K100M 9	0.996	10.326	0.969	0.156	0.968	23.606	0.987	0.761	15.623	0.979	0.046
E10FP 2	0.996	11.393	0.980	0.187	0.983	26.297	0.991	0.664	20.350	0.989	0.054
E10FP 9	0.937	15.397	0.998	0.425	0.983	36.938	0.990	0.392	43.513	0.997	0.113
KSR 2	0.989	11.920	0.973	0.202	0.976	27.493	0.985	0.673	20.987	0.982	0.057
KSR 3	0.784	16.026	0.997	0.825	0.879	39.806	0.927	0.202	61.979	0.981	0.171
KSR 9	0.944	13.861	0.999	0.318	0.987	33.152	0.991	0.411	37.952	0.996	0.085
pH 6.8											
K4M 2	0.997	10.986	0.972	0.173	0.975	25.181	0.989	0.733	17.455	0.982	0.050
K4M 3	0.948	16.217	0.968	0.561	0.983	39.362	0.989	0.302	53.009	0.968	0.144
K4M 9	0.990	12.811	0.990	0.237	0.996	29.837	0.997	0.591	25.898	0.996	0.066
K100M 2	0.998	7.769	0.985	0.104	0.974	17.638	0.994	0.836	10.336	0.990	0.031
K100M 3	0.949	15.577	0.991	0.412	0.985	37.135	0.988	0.430	41.332	0.995	0.109
K100M 9	0.995	10.075	0.969	0.149	0.967	22.928	0.988	0.812	13.960	0.978	0.044
E10FP 2	0.994	11.092	0.982	0.179	0.984	25.538	0.992	0.689	19.001	0.989	0.051
E10FP 9	0.935	14.844	0.999	0.381	0.982	35.546	0.988	0.406	41.007	0.997	0.101
KSR 2	0.992	11.578	0.983	0.191	0.984	26.576	0.992	0.719	18.845	0.990	0.055
KSR 3	0.797	15.700	0.985	0.610	0.889	38.587	0.925	0.272	54.335	0.980	0.156
KSR 9	0.959	13.485	0.999	0.291	0.993	32.119	0.995	0.433	35.555	0.998	0.078

to identify the release mechanism of drug. Savaşer *et al.* used the same application for drug release mechanism evaluation from diclofenac sodium containing HPMC matrices (Savaşer, Özkan, İşmer, 2005), Roni, Kibria and

Jalil (2009) applied equation for evaluation of controlled release alfuzosin tablets prepared with ethylcellulose and hydropropylmethylcellulose. El-Bagory *et al.* (2012) relate Korsmeyer-Peppas equation for release mechanism

TABLE V - Similarity Factor (f_2) Values of Tizanidine formulations

Comparison	f_2			Dissolution Profile
	0.1 N HCl	pH 4.5	pH 6.8	
K100M9 and K4M2	69.28	65.69	65.80	Similar
K100M9 and K4M3	20.42	20.96	22.44	Dissimilar
K100M9 and K4M9	43.66	42.05	42.22	Dissimilar
K100M9 and K100M2	47.22	48.88	49.19	Dissimilar
K100M9 and K100M3	23.73	24.33	26.37	Dissimilar
K100M9 and E10FP2	66.04	60.97	59.94	Similar
K100M9 and E10FP9	26.00	27.33	28.24	Dissimilar
K100M9 and KSR2	55.27	53.27	54.19	Similar
K100M9 and KSR3	16.57	20.68	21.93	Dissimilar
K100M9 and KSR9	28.13	33.48	34.45	Dissimilar

TABLE VI - Stability studies and shelf life of Controlled release tizanidine formulations

Study Period	Test	Methocel K4M			Methocel K100M			Ethocel 10 FP		Kollidon SR		
		K4M 2	K4M 3	K4M 9	K100M 2	K100M 3	K100M 9	E10FP 2	E10FP 9	KSR 2	KSR 3	KSR 9
0 Month	Hardness (kg)	4.81 ± 0.52	4.20 ± 0.38	4.69 ± 1.18	5.40 ± 0.68	4.30 ± 0.70	5.57 ± 0.76	12.92 ± 1.80	12.57 ± 2.30	5.01 ± 0.74	4.24 ± 0.88	4.20 ± 0.58
	Friability (%)	0.76	0.91	0.94	0.57	0.85	0.63	0.30	0.47	0.29	0.84	0.63
	*DT (Hrs)	4.82	4.18	4.30	6.87	4.95	6.58	5.72	5.13	7.52	4.25	6.27
	Dissolution (%)	98.32	97.98	97.56	82.32	98.15	99.52	98.21	98.65	99.59	98.29	98.53
	Assay (%)	98.76	97.25	98.05	98.37	97.82	98.87	98.79	98.24	98.39	96.51	97.27
	1 Month	Hardness (kg)	4.79 ± 0.61	4.15 ± 0.24	4.62 ± 0.94	5.36 ± 0.83	4.21 ± 0.78	5.55 ± 0.39	12.73 ± 1.75	12.49 ± 2.00	4.98 ± 0.83	4.20 ± 0.95
Friability (%)		0.77	0.91	0.94	0.68	0.9	0.65	0.38	0.54	0.35	0.87	0.69
*DT (Hrs)		4.78	4.04	4.25	6.82	4.78	6.51	5.64	5.03	7.18	4.03	5.93
Dissolution (%)		97.64	97.56	97.19	81.97	97.88	99.07	97.79	97.55	98.41	96.16	97.54
Assay (%)		98.53	96.79	97.64	98.08	97.34	98.51	98.32	97.86	98.17	95.87	96.82
3 Month		Hardness (kg)	4.76 ± 0.64	4.04 ± 0.73	4.58 ± 0.96	5.21 ± 0.88	4.13 ± 0.93	5.45 ± 0.86	12.45 ± 1.14	12.34 ± 2.20	4.96 ± 0.61	4.18 ± 0.56
	Friability (%)	0.81	0.93	0.97	0.63	0.94	0.69	0.42	0.63	0.42	0.93	0.78
	*DT (Hrs)	4.63	3.97	4.13	6.71	4.71	6.48	5.48	4.97	6.43	3.93	5.67
	Dissolution (%)	97.39	96.42	96.7	80.63	96.94	98.83	97.00	96.13	97.57	95.04	95.73
	Assay (%)	97.96	96.24	97.11	97.64	96.82	98.16	97.87	97.02	97.4	95.29	95.99
	6 Month	Hardness (kg)	4.68 ± 0.86	3.93 ± 0.65	4.53 ± 1.02	5.14 ± 0.93	4.01 ± 0.82	5.36 ± 0.54	12.30 ± 1.11	12.11 ± 1.64	4.83 ± 0.56	4.00 ± 0.98
Friability (%)		0.87	0.97	0.99	0.69	0.97	0.74	0.55	0.78	0.59	1.08	0.85
*DT (Hrs)		4.58	3.85	4.05	6.59	4.63	6.36	5.37	4.86	6.07	3.72	5.10
Dissolution (%)		96.84	95.47	95.93	80.28	96.1	98.54	96.13	94.33	96.52	91.76	93.33
Assay (%)		97.84	95.73	96.85	97.38	96.51	98.02	97.46	96.77	96.83	94.83	95.00
Shelf Life (Months)		26.680	18.896	21.712	29.450	19.775	30.347	24.172	17.053	21.900	13.551	16.209

*DT = Disintegration time

determination of theophylline matrix tablets prepared with Kollidon SR, Carnuba wax and ethylcellulose.

Model independent method

Formulation K100M9 was considered as reference and f_2 similarity test was performed for selected formulations. Dissolution profile was found to be similar for K4M2, E10FP2 and KSR2 (Table V). Comparison of dissolution profiles of drug through similarity test (f_2) was used in other research studies for SR preparations (Dash *et al.*, 2010; Shoaib *et al.*, 2010; Salústio *et al.*, 2011).

Response surface analysis

Response surface models of effect of HPMC K4M and Avicel pH 101 are shown in Figure 5a, 5b and 5c. The 3D plots clearly showed that polymer had greater influence on response as compared to Avicel pH 101. Disintegration time increased drastically with an increase in polymer concentration while Avicel has lesser effect in enhancing disintegration time as compare to HPMC K4M. Increase in HPMC K4M ratio in the formulation caused a steep decrease in the release in 2 hours and Avicel seems to produce not a very significant impact on matrix release.

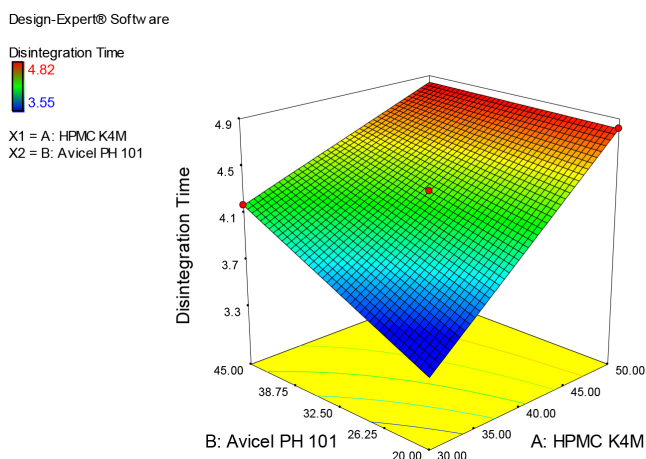


FIGURE 5a - Response surface model (RSM) showing effect of independent variables HPMC K4M and Avicel pH 101 on Disintegration time.

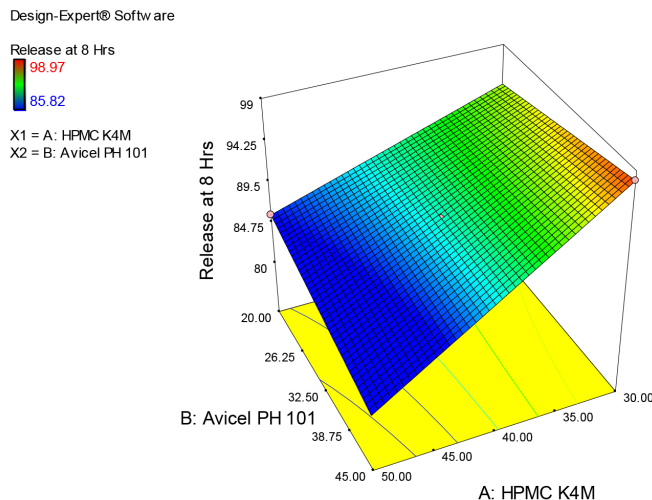


FIGURE 5c - Response surface model (RSM) showing effect of independent variables HPMC K4M and Avicel pH 101 on Drug release in 8 hours.

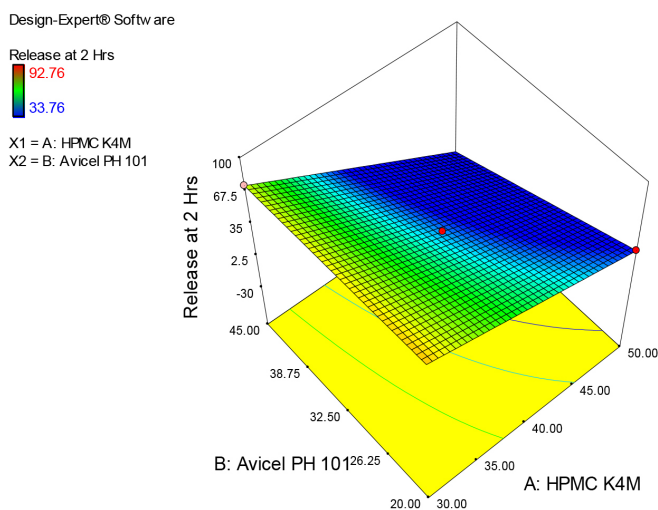


FIGURE 5b - Response surface model (RSM) showing effect of independent variables HPMC K4M and Avicel pH 101 on responses Drug release in 2 hours.

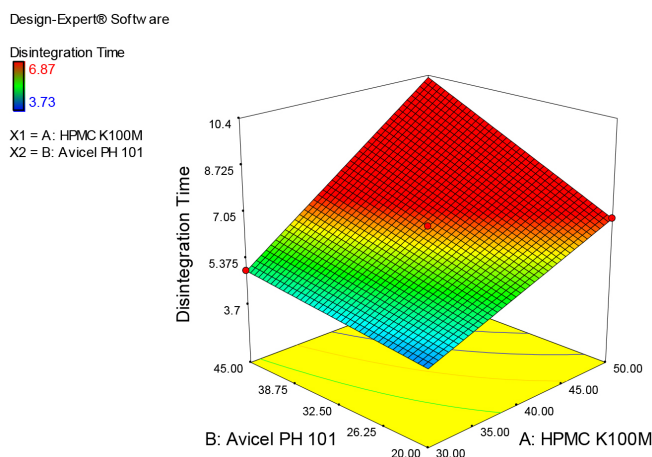


FIGURE 6a - Response surface model (RSM) showing effect of independent variables HPMC K100M and Avicel pH 101 on Disintegration time.

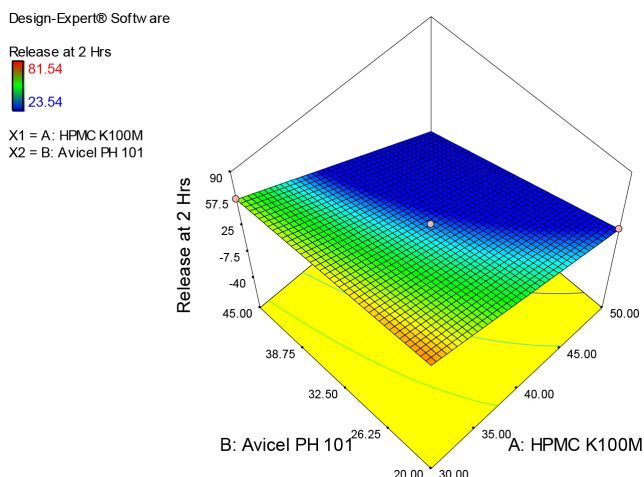


FIGURE 6b - Response surface model (RSM) showing effect of independent variables HPMC K100M and Avicel pH 101 on Drug release in 2 hours.

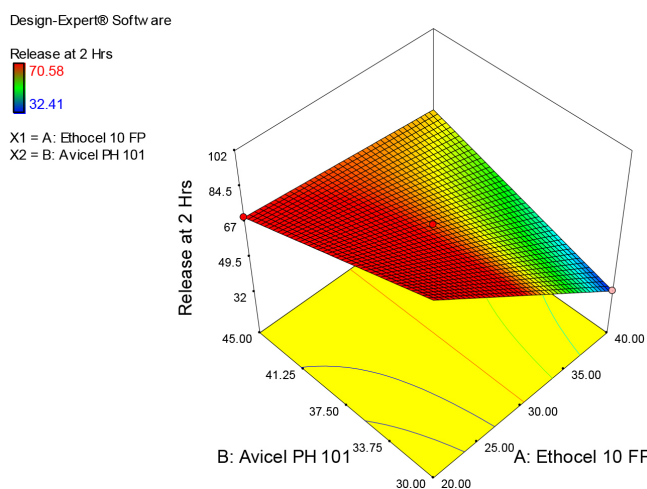


FIGURE 7b - Response surface model (RSM) showing effect of independent variables Ethocel 10FP and Avicel pH 101 on Drug release in 2 hours.

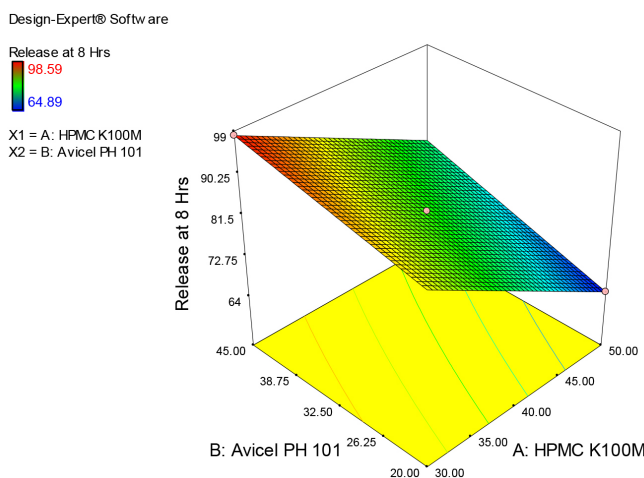


FIGURE 6c - Response surface model (RSM) showing effect of independent variables HPMC K100M and Avicel pH 101 on Drug release in 8 hours.

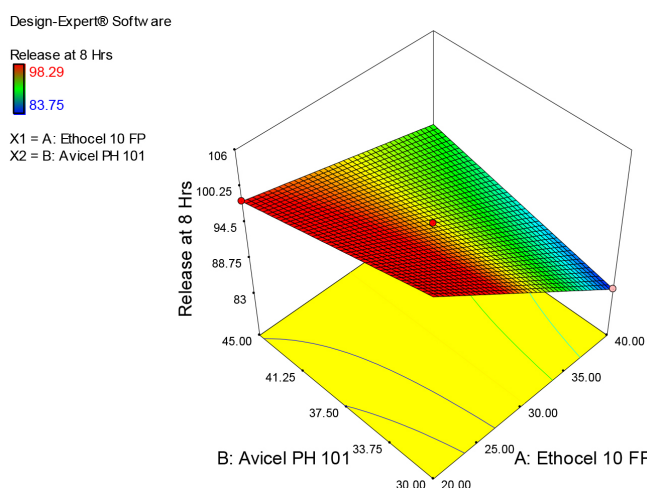


FIGURE 7c - Response surface model (RSM) showing effect of independent variables Ethocel 10FP and Avicel pH 101 on Drug release in 8 hours.

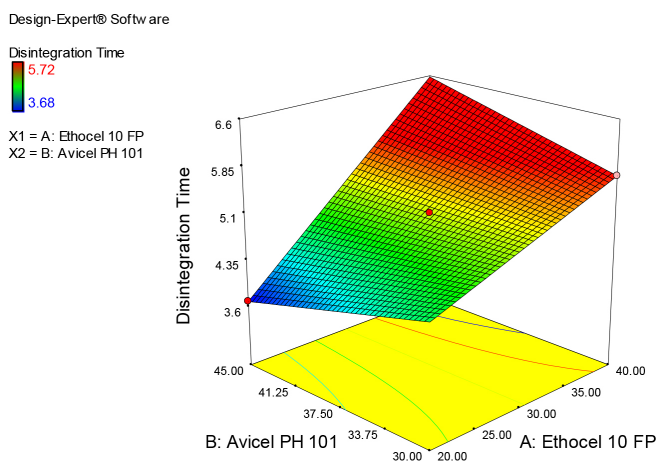


FIGURE 7a - Response surface model (RSM) showing effect of independent variables Ethocel 10FP and Avicel pH 101 on Disintegration time.

At 8 hours, no prominent effect on release can be seen with HPMC K4M change and a near to linear response found with change in Avicel pH101. Similar responses were observed with HPMC K100M for disintegration time and release at 2 hours while release at 8 hours has been significantly influenced with increase in HPMC K100M as presented in Figure 6a, 6b and 6c. RSM for Ethocel 10FP effect on responses were shown in Figure 7. It can be clearly seen that increase in concentration of polymer caused an increase in disintegration time and Avicel did not produce any significant change in disintegration time (Figure 7a, 7b and 7c). There was a declining response in release of the drug at 2 hours and 8 hours with an increase

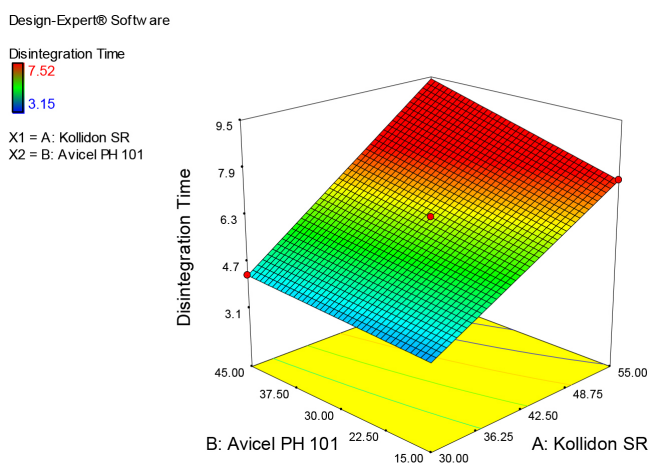


FIGURE 8a - Response surface model (RSM) showing effect of independent variables Kollidon SR and Avicel pH 101 on Disintegration time.

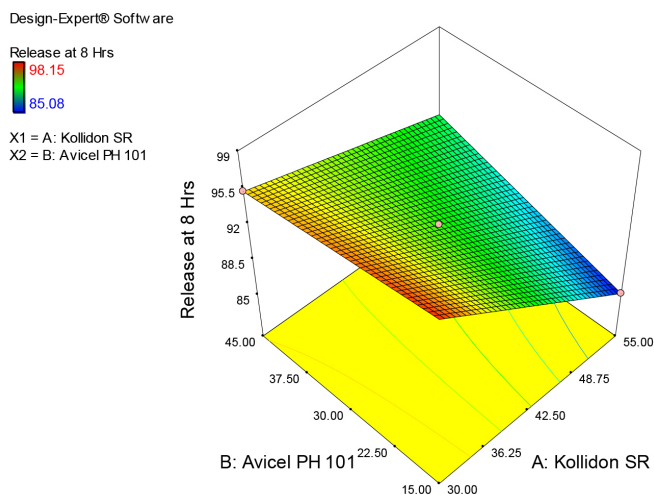


FIGURE 8c - Response surface model (RSM) showing effect of independent variables Kollidon SR and Avicel pH 101 on Drug release in 8 hours.

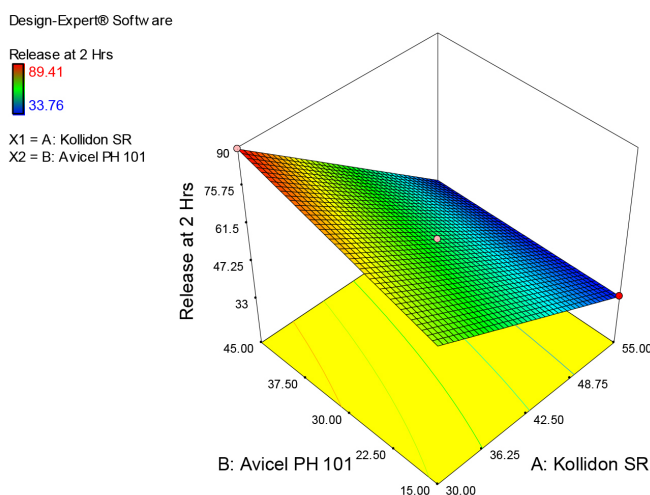


FIGURE 8b - Response surface model (RSM) showing effect of independent variables Kollidon SR and Avicel pH 101 on Drug release in 2 hours.

in Ethocel 10FP proportion in the formulation (Figure 7b and 7c). Kollidon SR produced expected response of increase in disintegration time with increase in polymer concentration. An inverse relationship found between Kollidon SR proportion and release at 2 and 8 hours (Figure 8b and 8c). Bose, Wong and Singh (2012) found similar type of effect of HPMC K100M on Itopride SR matrix tablet release.

Stability evaluation

The formulations showing disintegration time greater than 4 hours were subjected to accelerated stability studies for 0, 1, 3 and 6 months following ICH guidelines

(ICH, 2003). Formulation K4M2, K4M3, K4M9, K100M2, K100M3, K100M9, E10FP2, E10FP9, KSR2, KSR3 and KSR9 were evaluated for quality attribute, shelf life were calculated using software R-Gui version 2.15.2 (stab) and results were given in Table VI. It was found that all formulations were within the acceptable limits for physical and chemical parameter but formulations K4M3 and KSR3 were failed for disintegration time and friability. It was found that formulations K4M2, K100M2, K100M9 (highest shelf life: 30.347 months) and E10FP2 had the shelf life of more than 24 months and were considered as stable formulations.

CONCLUSION

Tizanidine hydrochloride formulations with controlled release characteristics were prepared by using HPMC K4M and K100M, Ethocel 10FP and Kollidon SR.

It was observed that the formulations with polymer contents lesser than 40% remained ineffective in controlling the release of drug however, satisfactory release profiles were obtained with HPMC K4M (K4M2) 50%, K100M (K100M9) 40%, Ethocel 10FP (E10FP2) 40% and Kollidon SR (KSR2) 55%. Formulation K100M9 with HPMC K100M showed highest shelf life of 30.347 months. Therefore, it can be concluded that controlled release tizanidine hydrochloride tablets can effectively be prepared by using these polymers through direct compression method. Moreover, formulation K100M9 was found to be the best formulation for controlled release tizanidine.

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